

An aerial photograph of a forested hillside. The trees are in various stages of autumn, with many showing bright yellow and orange foliage, while others remain green. The terrain is uneven, with some rocky outcrops visible. The overall scene is a dense forest with a mix of colors and textures.

# Functional Genomics of *Populus* Growth & Development

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# Outline

- Environmental regulation of growth, dormancy, flowering
  - Regulatory genes and differentiating functions of paralogs
  - Multi-tissue time series transcriptomics to gene regulatory networks
- Wood-associated protein-protein and protein-DNA interactions
  - Have we identified new regulators of wood formation?

## Phenology is closely matched to local climates

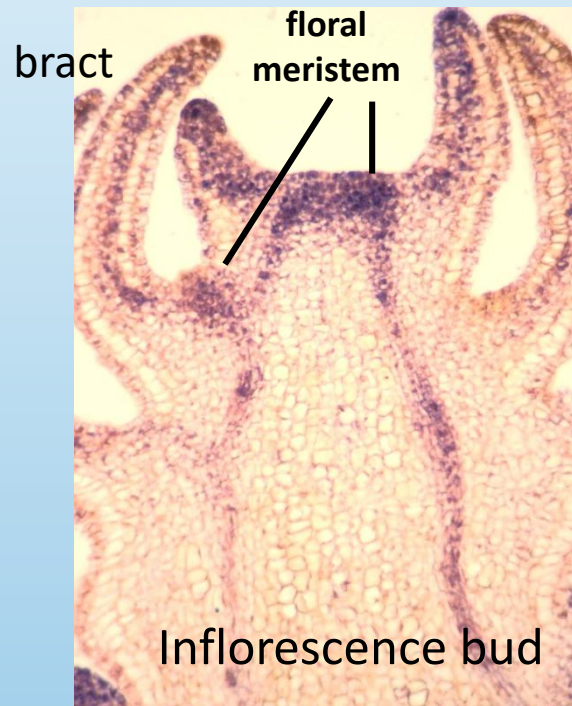
- Common garden studies
  - Latitudinal clines
  - Genetic differentiation between populations
- Ecotypes –controlled environment studies
  - Different critical daylengths for bud set & dormancy induction
  - Phytochrome: night breaks of white or red light disrupt SD response



- **Dormancy:** a meristem is insensitive to growth promoting conditions until it is released from dormancy by an environmental cue
- **Depth of Dormancy:** refers to the quantitative nature of this phase
- “Classic terms”
  - Ecodormancy: meristem is quiescent, but will rapidly resume growth if the limiting environmental factor is altered
  - Endodormancy



- **Reproductive phenology:** Seasonal timing of the floral transition, anthesis and fruiting
  - In temperate zones, flowering is indirect



# The environmental signals regulating vegetative and flowering phenology in temperate zones are generally the same

Environmental Factor	Reliability in temperate zones
Photoperiod	High
Prolonged chilling period	High
Ambient temperature	Moderate
Light intensity	Moderate
Water availability	Moderate
Nutrient availability	Low
Light quality	Low

*Adapted from Bernier and Perilleux (2005) A physiological overview of the genetics of flowering time control. Plant Biotechnol J 3 (1): 3-16*

**Less predictable factors modulate the effects of the primary signals and sometimes can substitute for the primary signal**

# Poplar Phenology

Growth cessation	Bud set
<b>Photoperiod</b> Temperature Light quality/quantity Water availability <b>Nutrient availability</b>	

<b>Dormancy induction</b>	<b>Dormancy release</b>
Photoperiod Temperature	Chilling sum Photoperiod

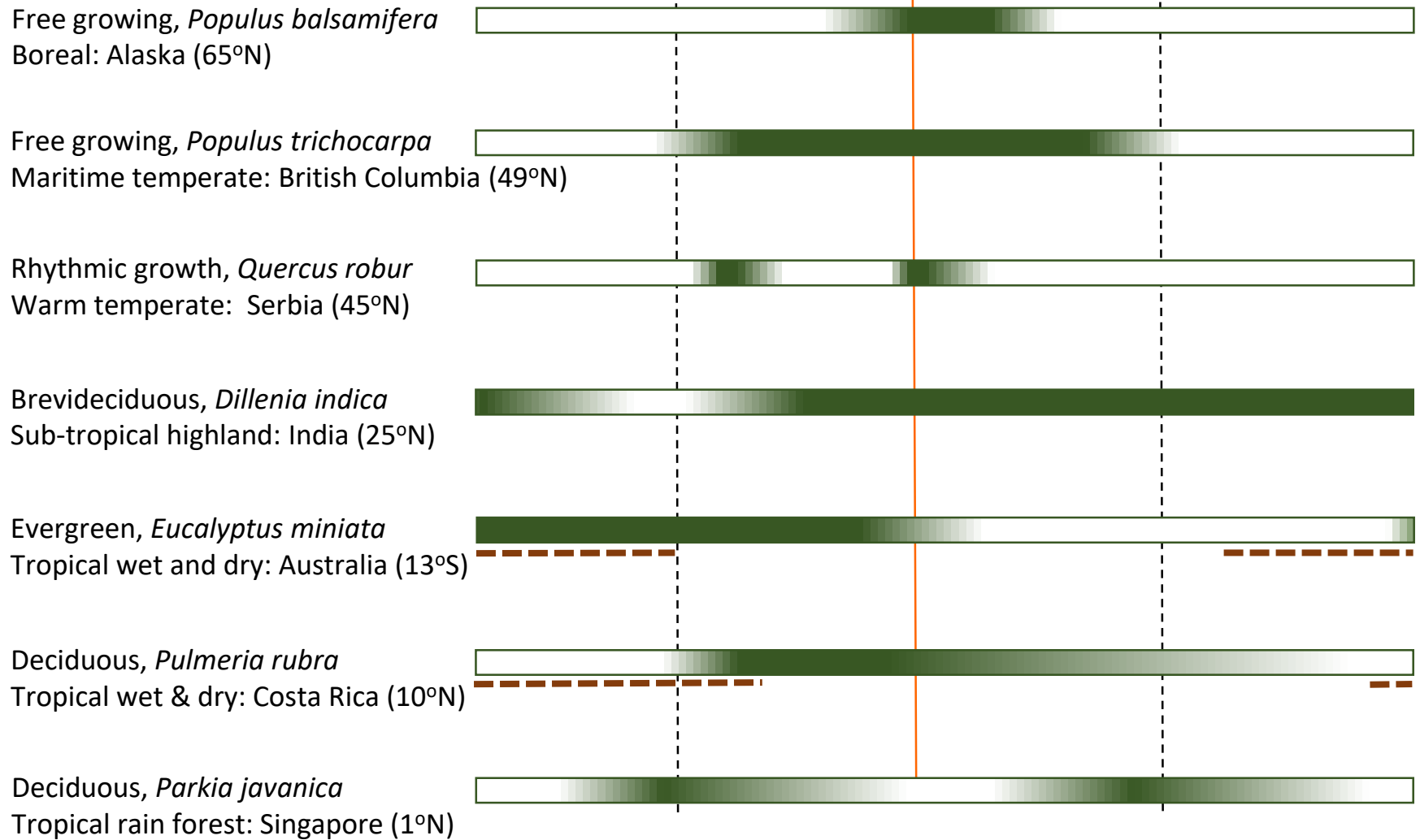
Leaf senescence/N storage	Leaf Drop
<b>Temperature</b> Photoperiod, light quality/quantity	

Cold acclimation	Cold deacclimation
Photoperiod Temperature	Heat sum Photoperiod

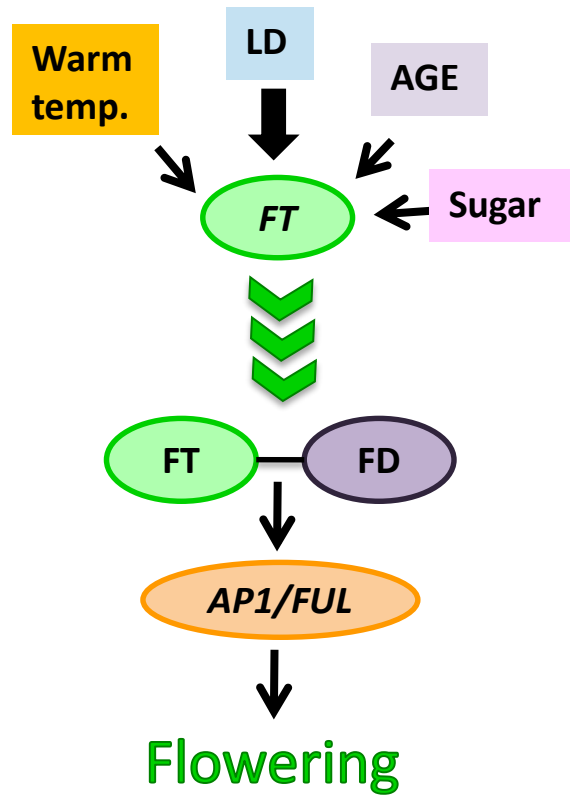
Growth resumption	N recycled	Bud flush
Heat sum Photoperiod  Water/ <b>nutrient availability</b>		

# Tree annual growth patterns

Spring Equinox      Summer Solstice      Autumn Equinox

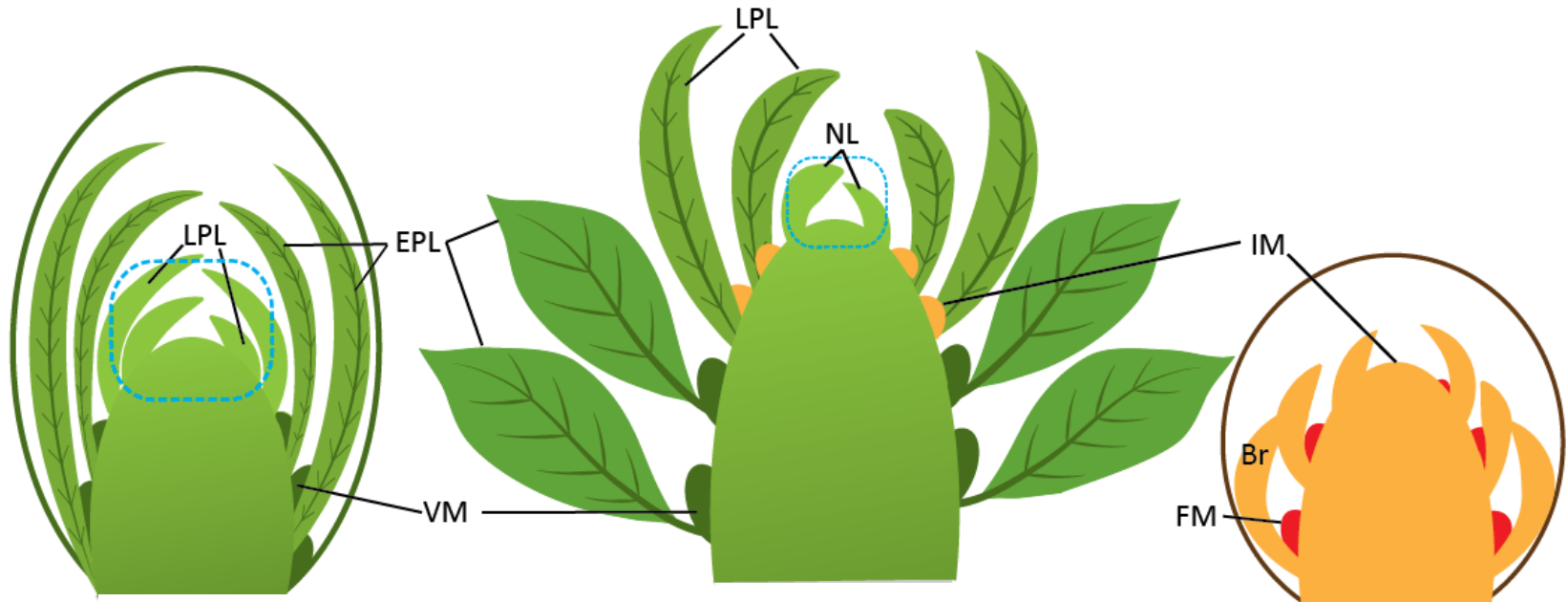






**FT is an integrator of environmental and internal signals regulating flowering time**

# Transition to flowering in *Populus*



Dormancy release

Bud flush

Inflorescence  
meristems form

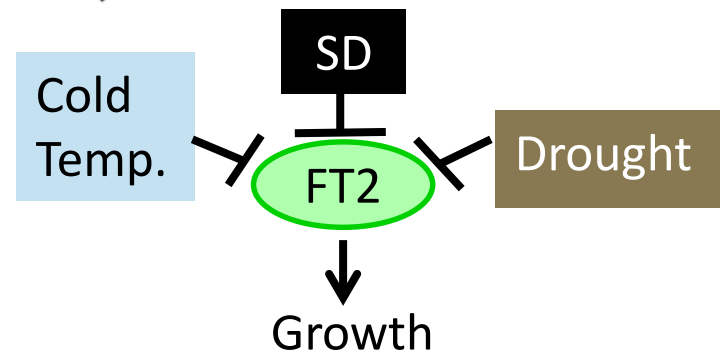
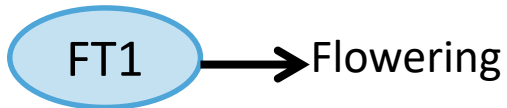
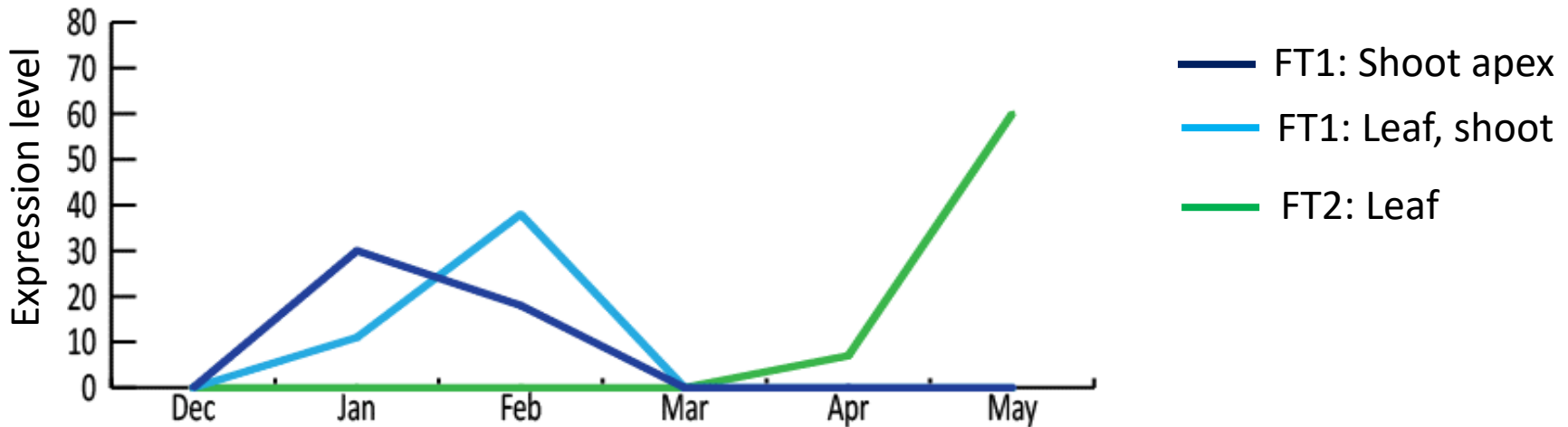
Floral meristems  
form

Meristems commit to flowering during a limited seasonal time

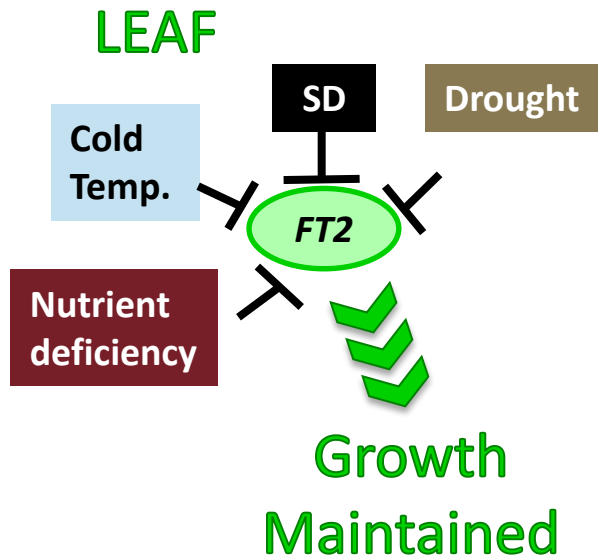
# FT1/FT2 seasonal expression in mature *P. deltoides*

Hsu et al. 2011 PNAS 108: 10756-61

Dormancy  
release  
.....  
Bud  
flush    IM  
form    FM  
form →



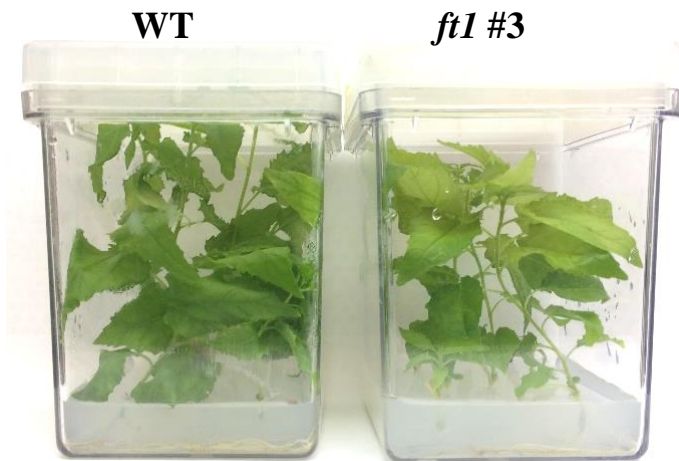
***Do FT1 and FT2 have distinct roles in vegetative phenology?***



CRISPR/Cas9-induced mutations in both *FT2* and *FT1*: reduced shoot elongation and terminal bud set in tissue culture under 16 hr daylengths



*FT1*-specific mutants appear WT

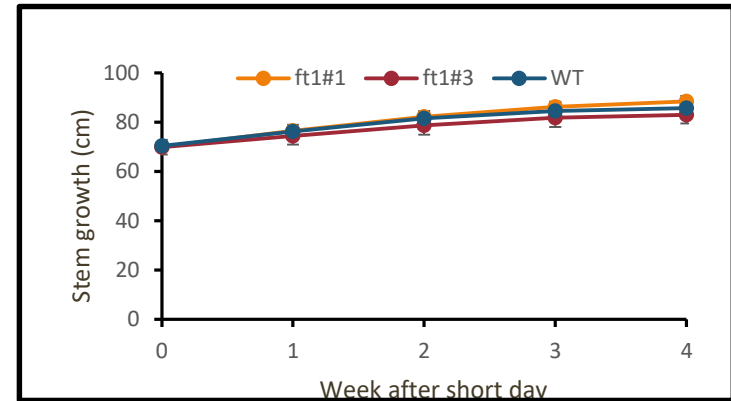


# *ft1* mutants have WT-like phenotype in both long and short daylengths

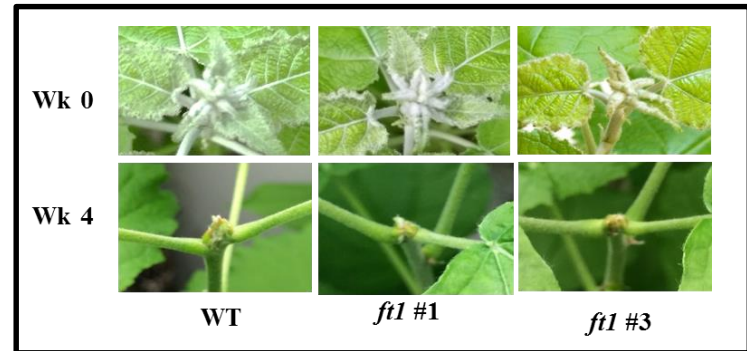
## Growth under long days



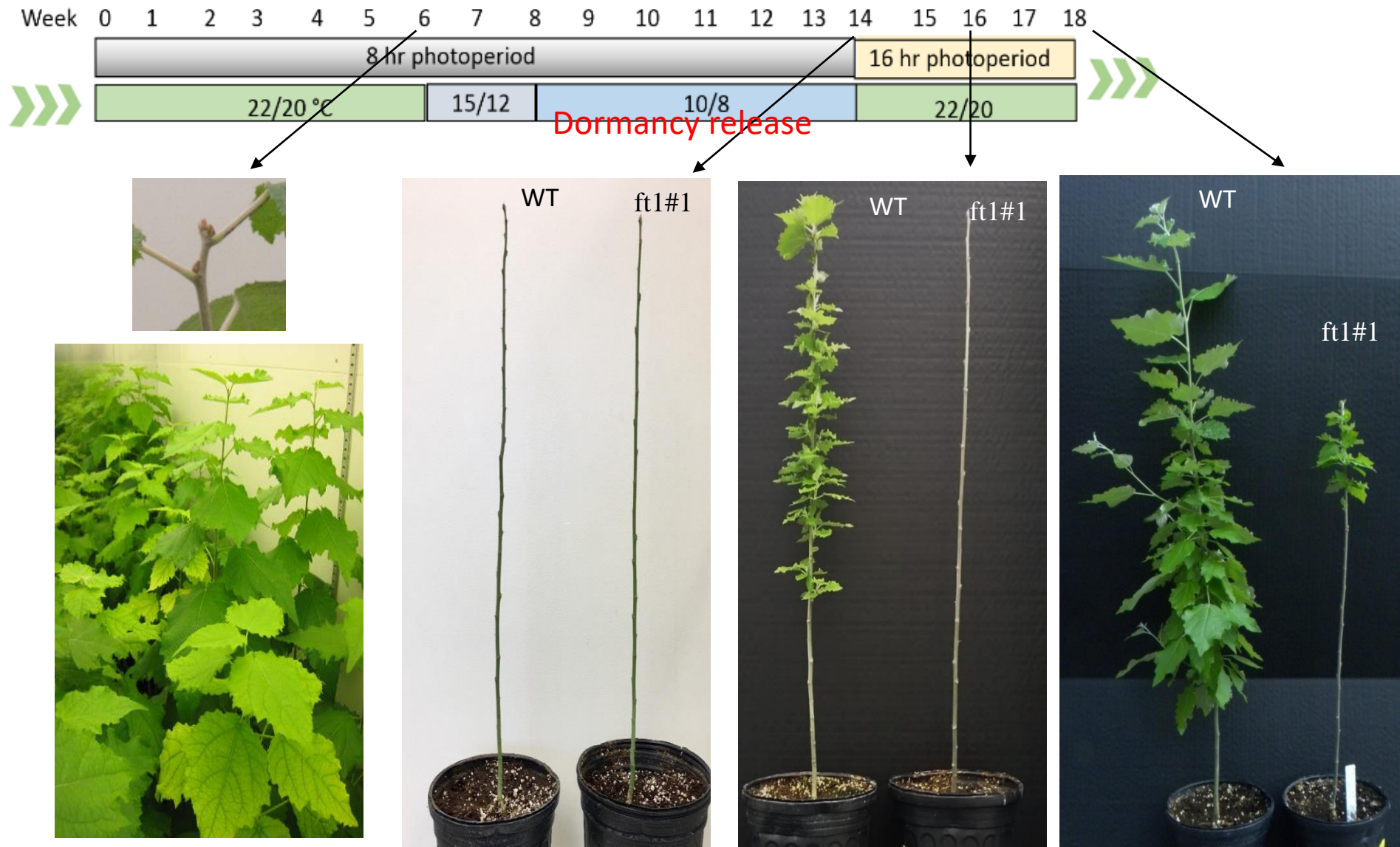
## Growth cessation under short days

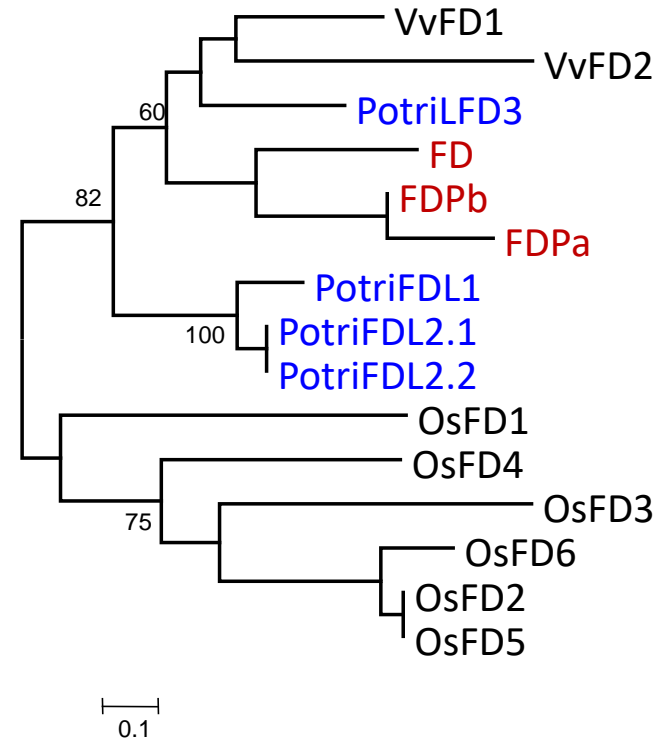
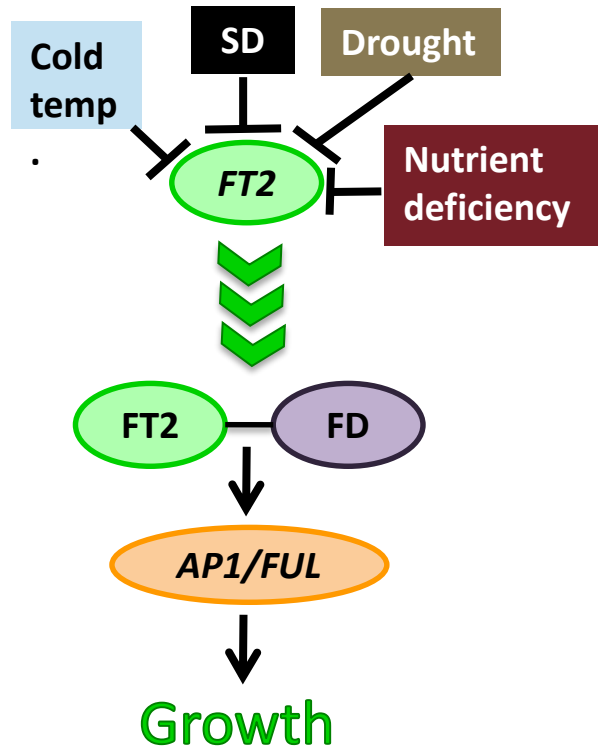


## Bud set



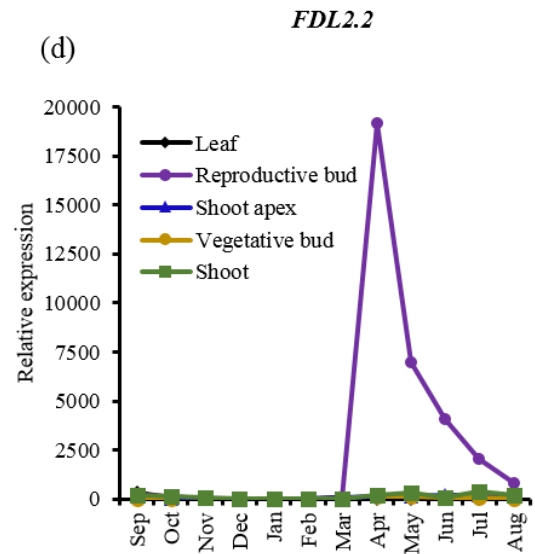
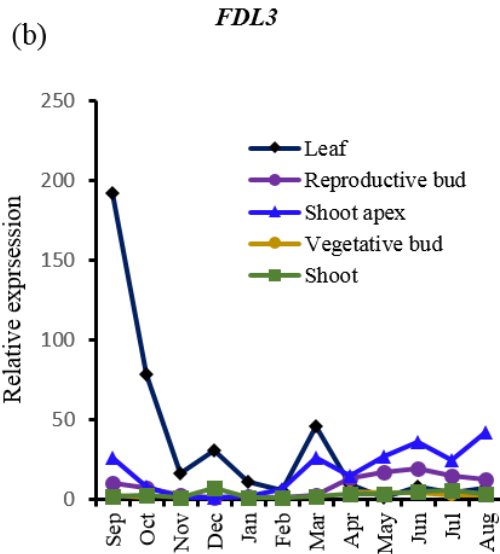
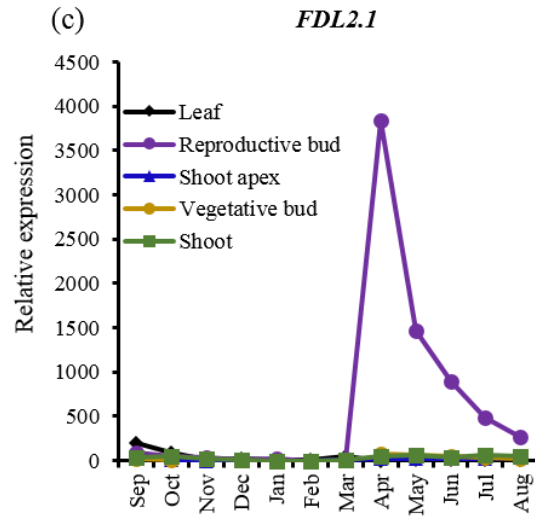
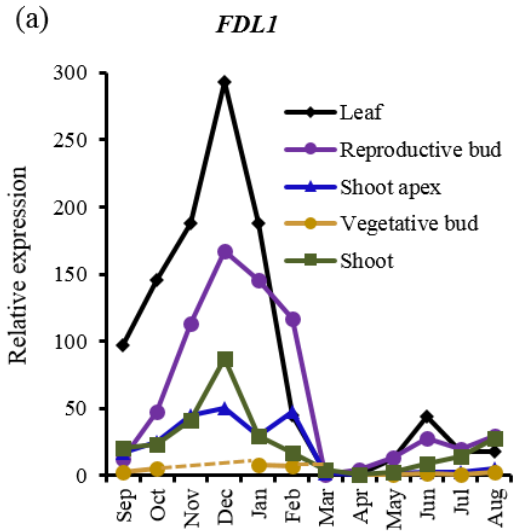
# Dormancy release is delayed in *ft1* mutants





FDL2.2: Parmentier-line and Coleman 2016

FDL1 and FDL2.1 :Tylewicz et al. 2015



- All *Populus* *FDLs* delay SD-induced bud set
- Only *FDL2.2* induced flowering under LDs
- *FDL2* and *FDL3* affect shoot development under long days
- Diverged in regulation and proteins have partial functional equivalency



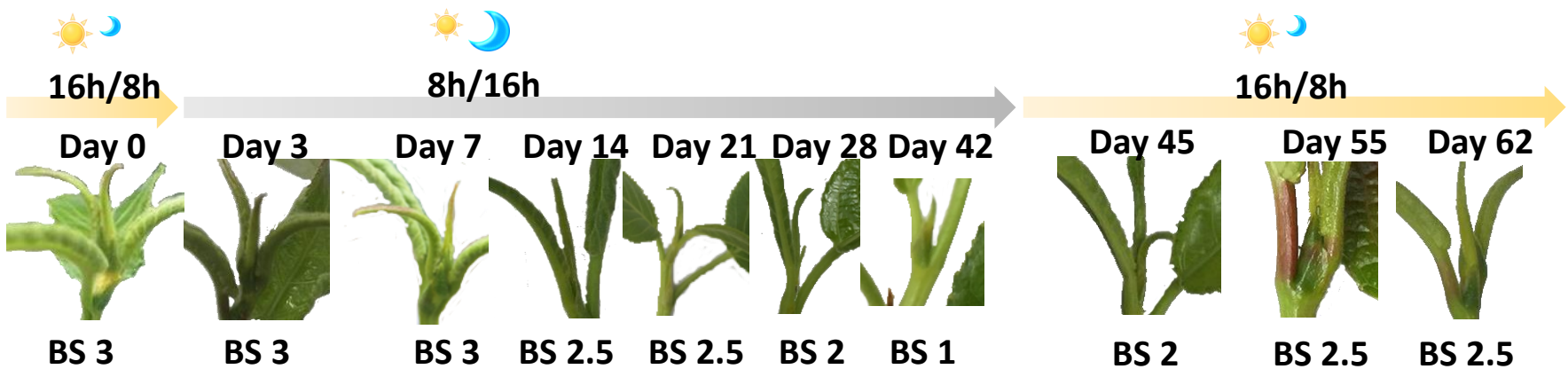
# Multi-tissue time series transcriptomes

- 540 *P. trichocarpa* Nisqually1
- Daylength
  - LD → SD → LD
- Nutrient
  - HN → LN → HN
- Tissues/organs:
  - Shoot apex
  - Leaf
  - Root
  - Cambial zone (daylength only)

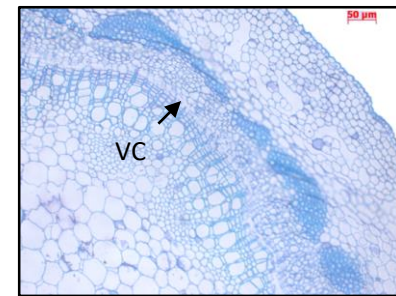
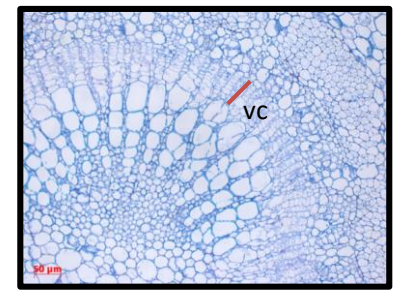
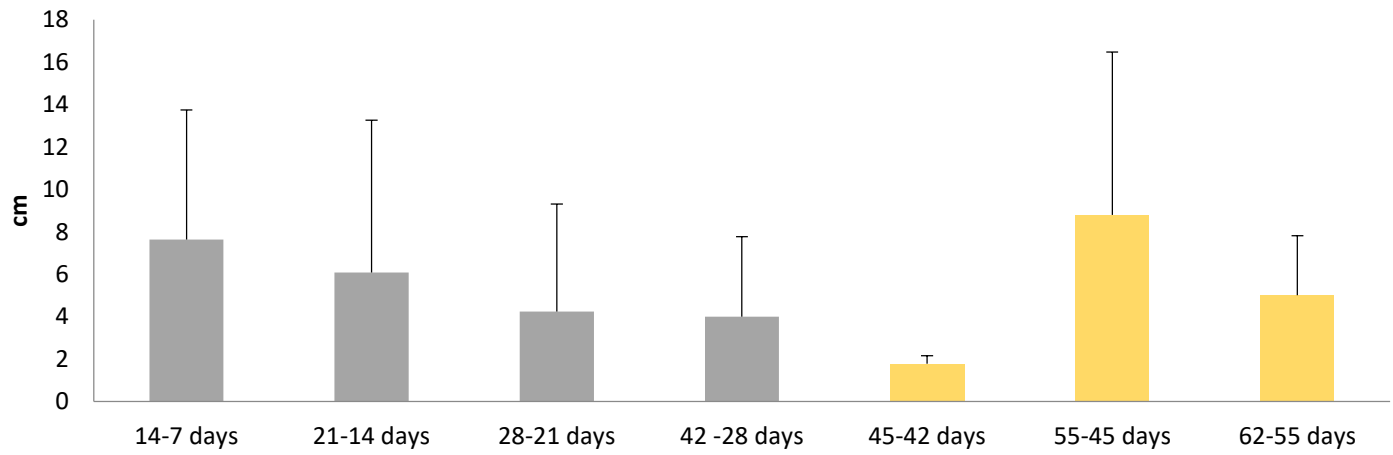


## Goals/Questions

- GRNs for organismal-level processes and phenotypes relatable to field conditions/natural populations
- Among different organs/tissues and environmental treatments:
  - To what extent are there common modules, transcriptional regulators (context dependent), paralogous modules/regulators etc.?
- Regulatory network context of adaptive variation



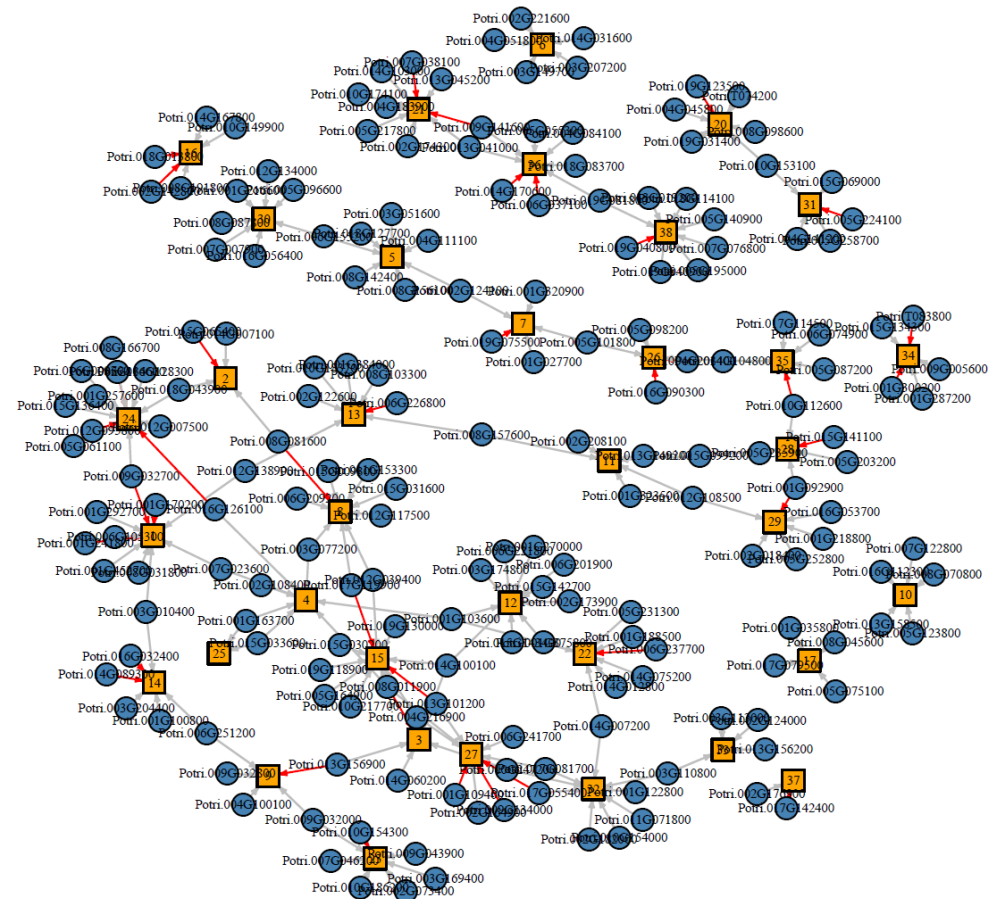
**Shoot elongation**



# Gene Regulatory Network (GRN) Prediction: Ensemble method

- Uses 5 different algorithms: ARACNE, Random Forest, Least angle regression, Partial correlation, Context likelihood relatedness (Redekar, N. et al. 2017)

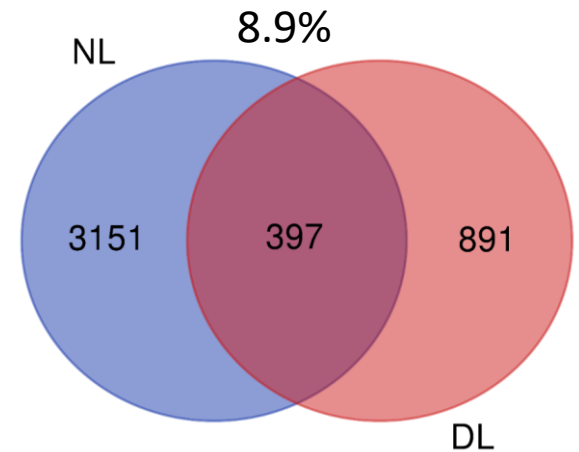
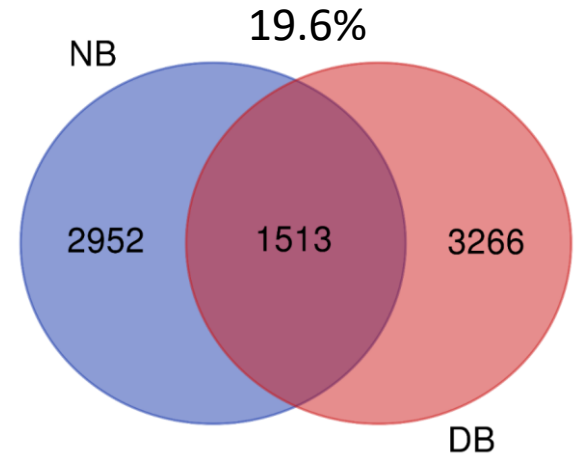
Transcriptional regulators predicted by at least 4 of the 5 algorithms



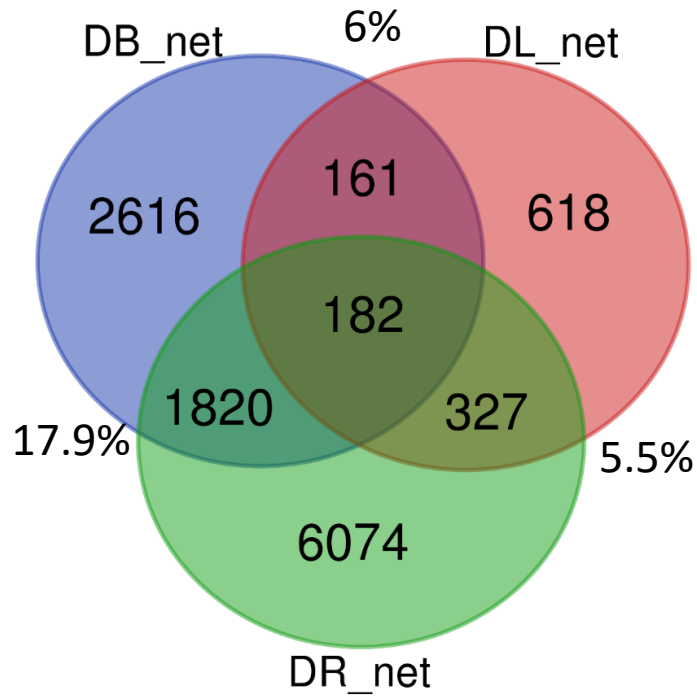
More unique genes per network, but also common genes between:

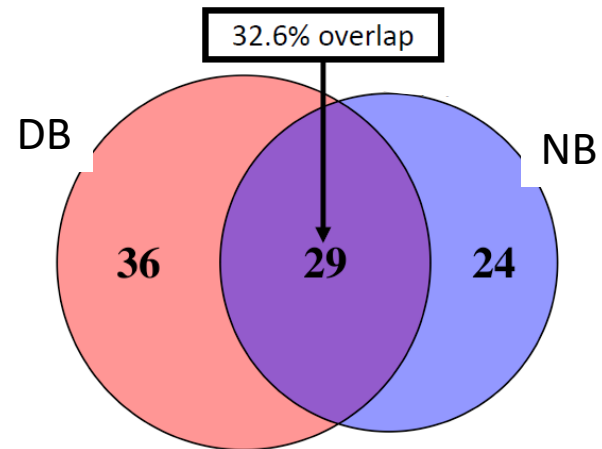
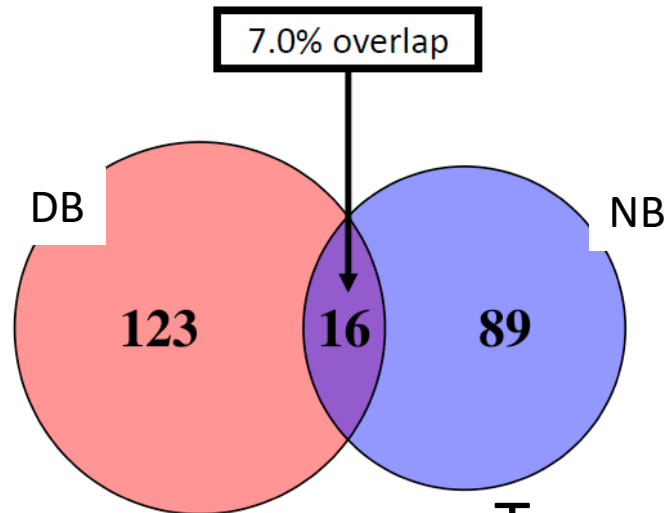
- Different organs in same treatment
- Same organ in different treatments

Same organ, nutrient & daylength

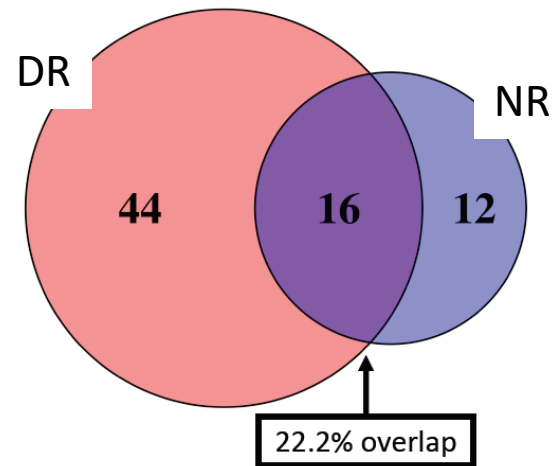
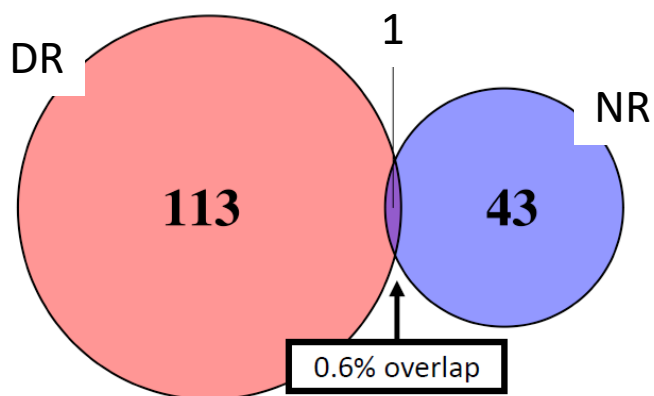
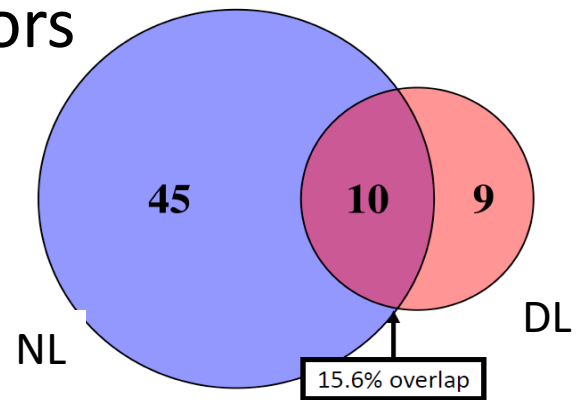
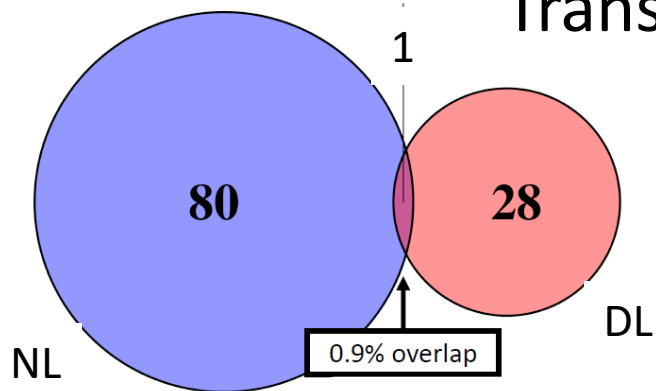


3 Daylength organs

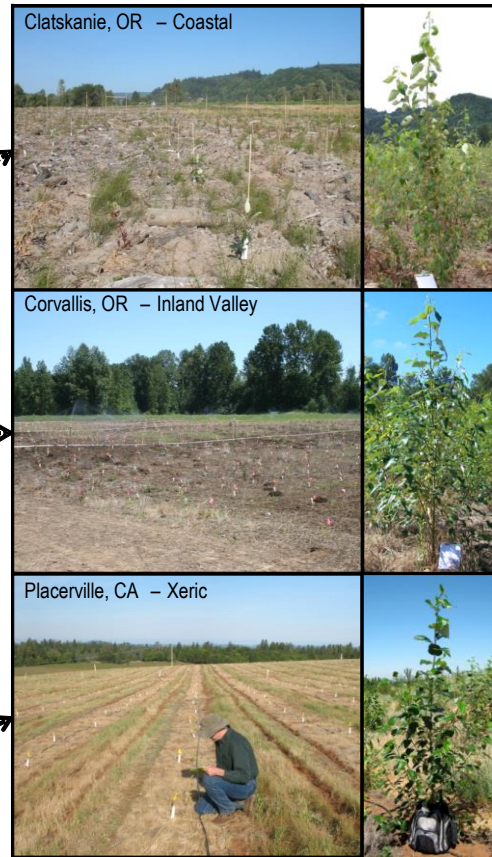
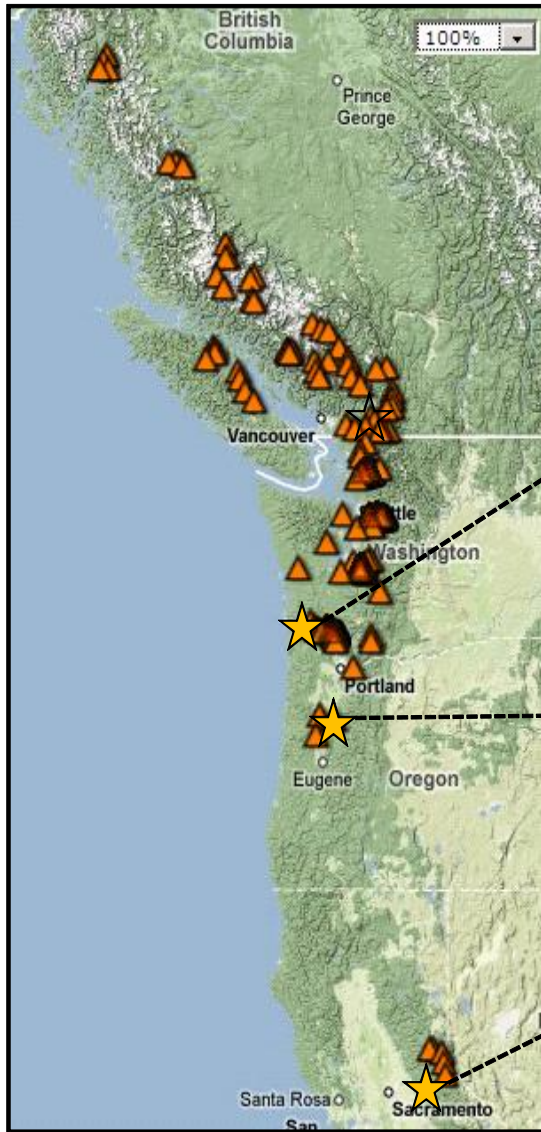




### Transcriptional regulators



# Regulatory context of adaptive variation



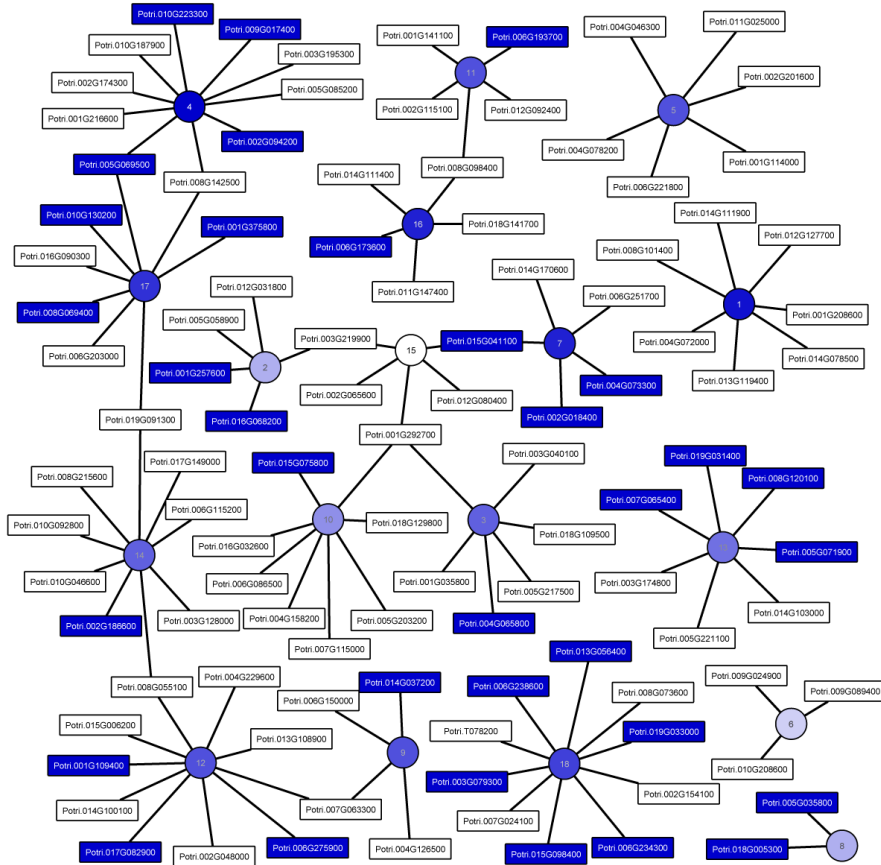
(Evans et al. 2014)

- GWAS traits

- Bud set
- Bud flush
- Height

- Selection scans

# Nutrient Bud Network - GWAS

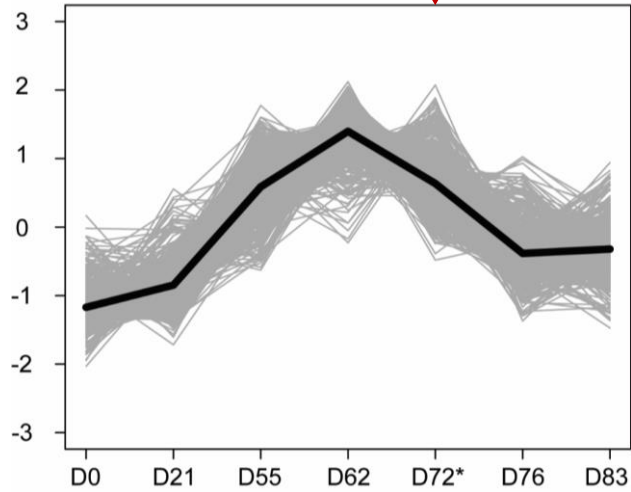


.19  .35

- 115 Transcriptional regulators
  - 33 GWAS
- 18 modules (4465 genes)
  - 96-404 genes/module
  - 1318 GWAS



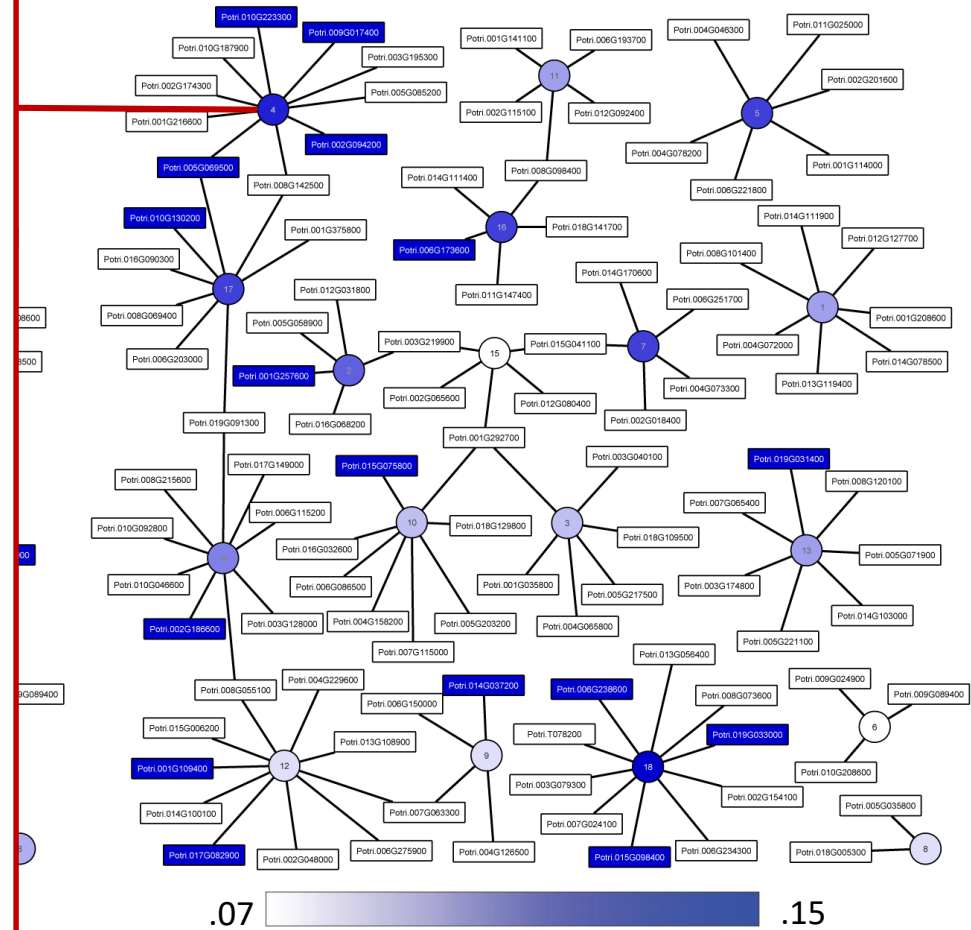
Fertilization



■ Median Module Expression Pattern  
 ■ Module Member Expression Pattern

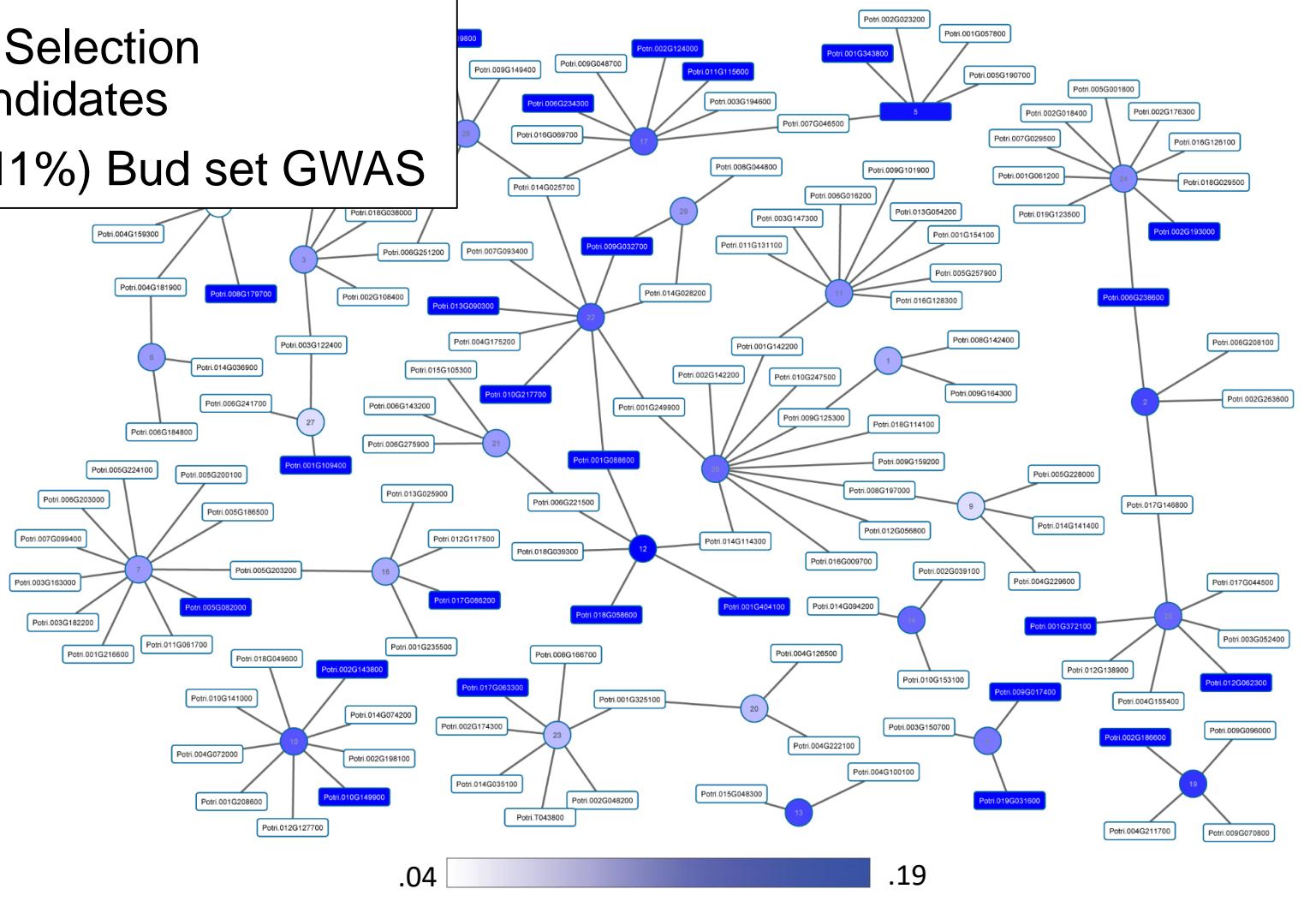
- Tale transcriptional regulator
  - Bud set associated gene
  - Selection candidate
  - Also regulator in the daylength bud network

### Nutrient Bud Network - BS-GWAS



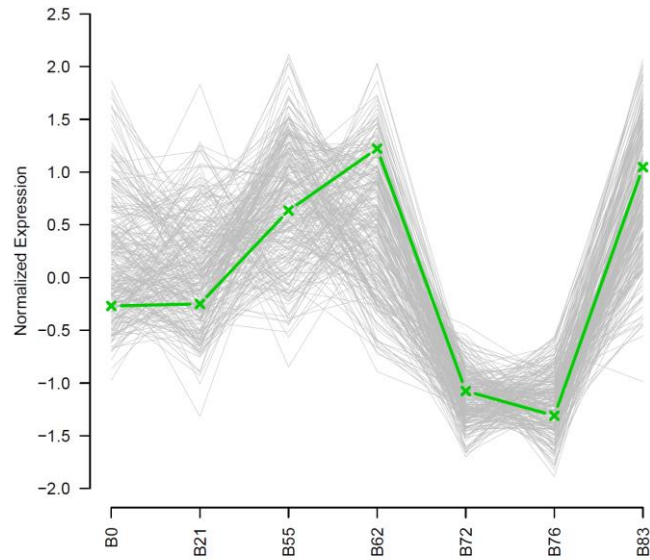
- 163 transcriptional regulators
- 27 Bud set GWAS
- 24 Selection candidates
- 529 (11%) Bud set GWAS

Day Length Bud – BS-  
GWAS



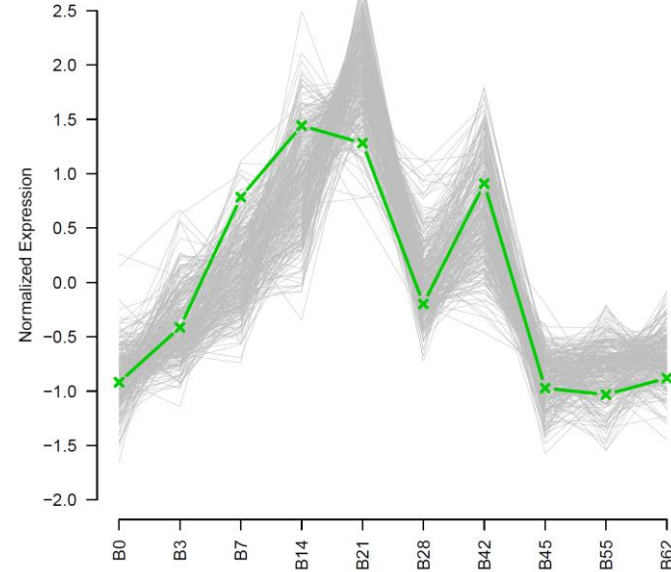
# FDL1 is a predicted regulator in nutrient and daylength bud networks

NB K7 FDL1



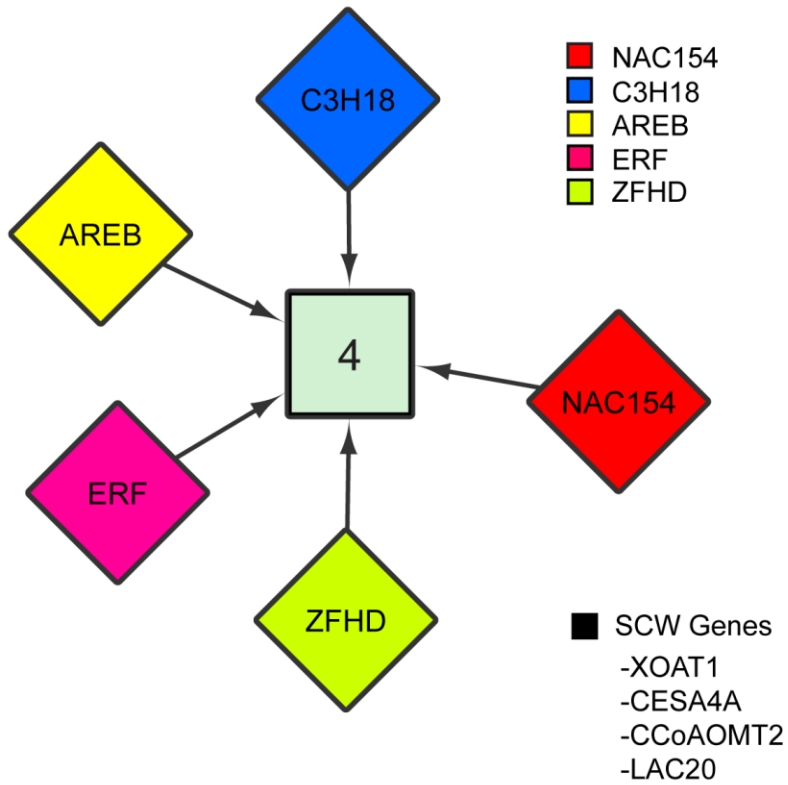
- Response to Chitin
- Regulation of JA signaling

DB K29 FDL1

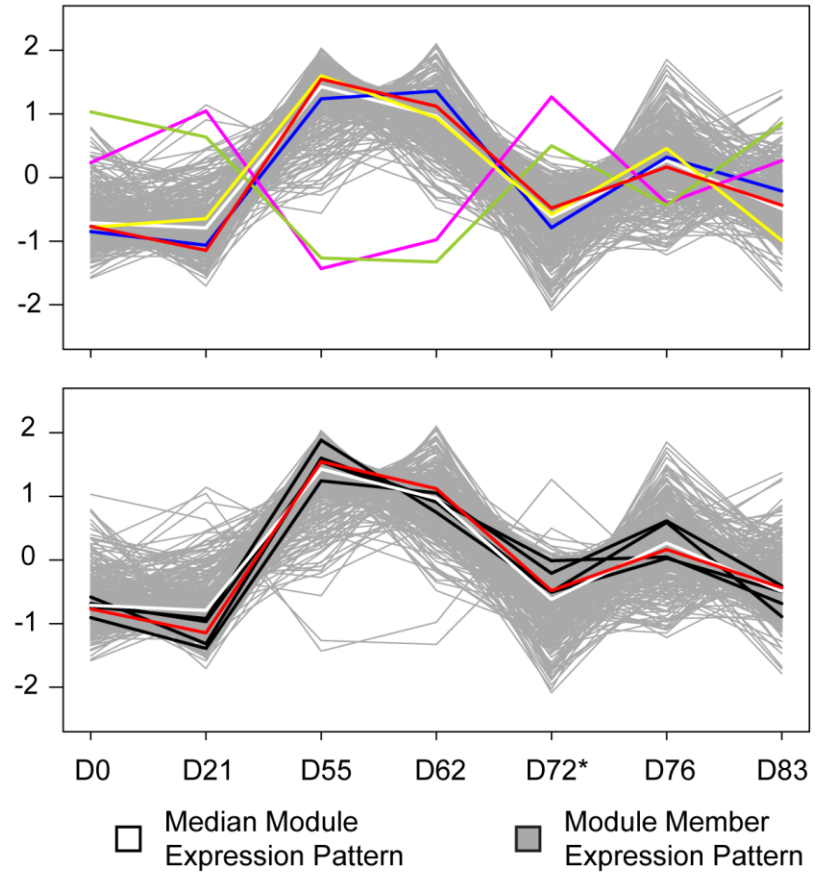


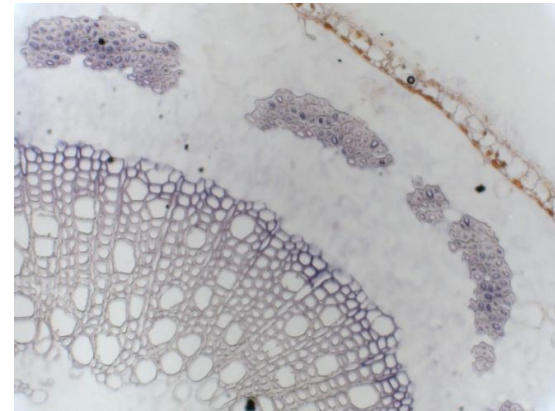
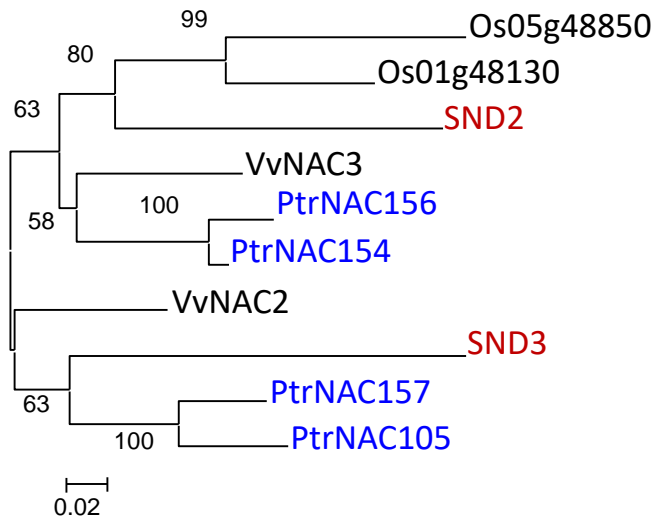
- Response to water stress
- Response to ABA

# Module from leaf nutrient response GRN



Module 4 Leaf Nutrient Deficiency Time Course





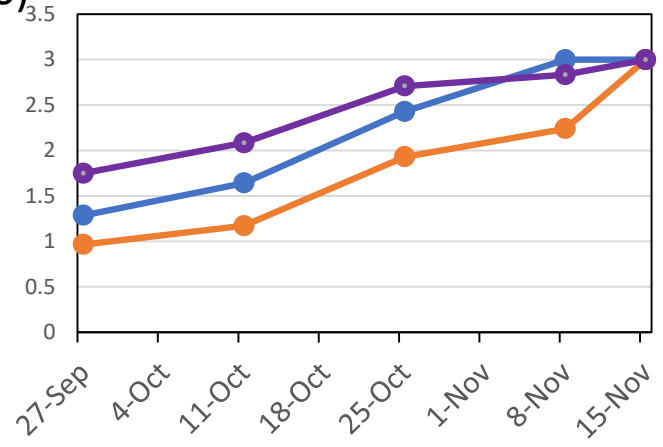
- *35S:NAC154* poplar (Grant et al. 2010; Jervis et al. 2015)
  - Reduced size
  - Elevated levels of arginine in stems
  
- *35S:PtrSND2-SRDx* poplar (Wang et al. 2013)
  - Reduced growth
  - Reduced secondary cell wall thickening

amiRNA-SND2 (targets paralogs NAC154 and NAC 156)

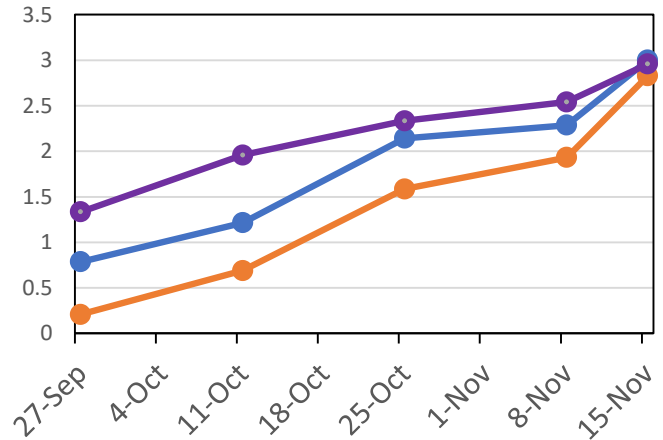


Field trial established Nov 2013, photo taken Dec 4, 2015

## Leaf senescence

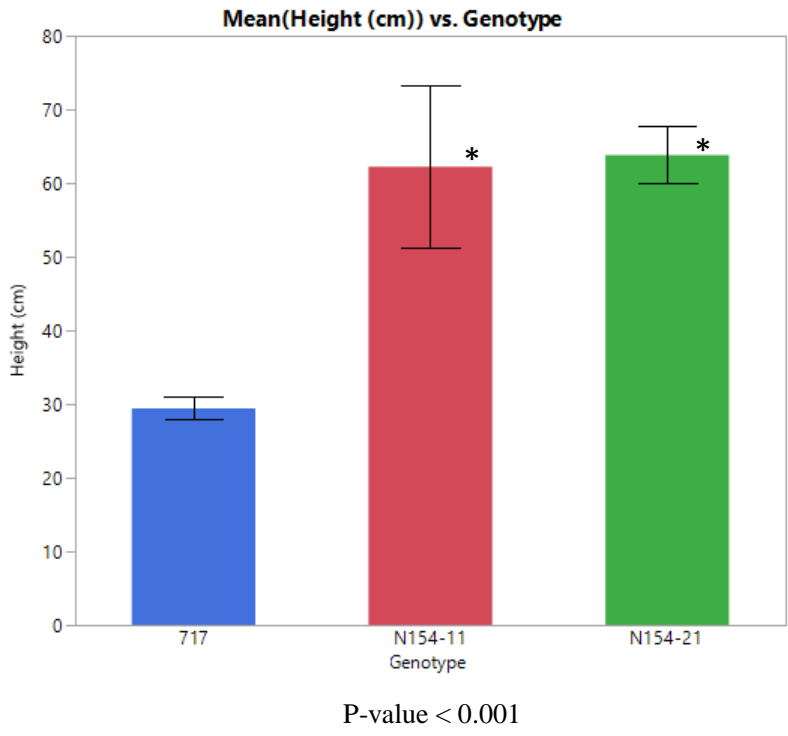


## Leaf drop

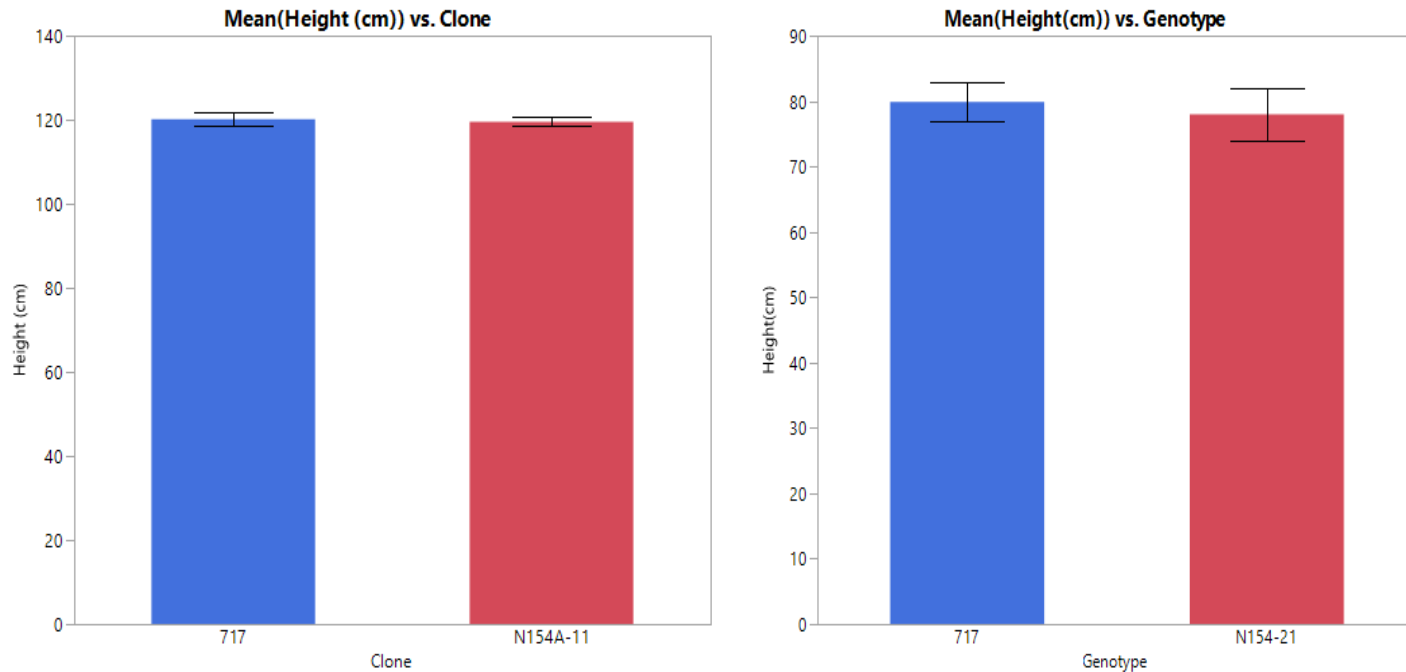


—●— WT —●— N154A —●— N154X

# N154-LN study



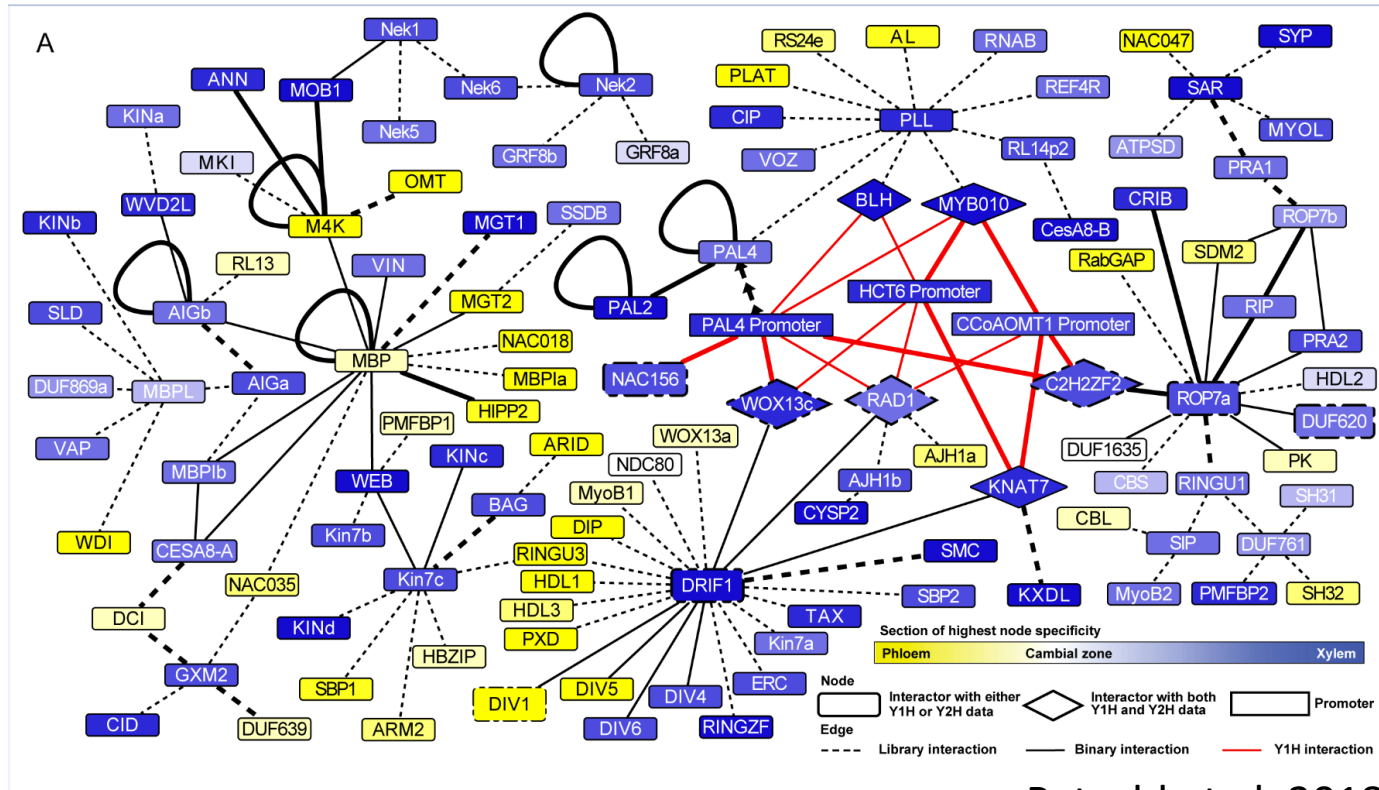
- In short days, amiRNA-SND2 transgenics cease growth and set bud the same time as WT
- After temperature lowered, leaf senescence and drop is slower in transgenics



Results consistent with presence of NAC154 in nutrient network, but not the daylength network



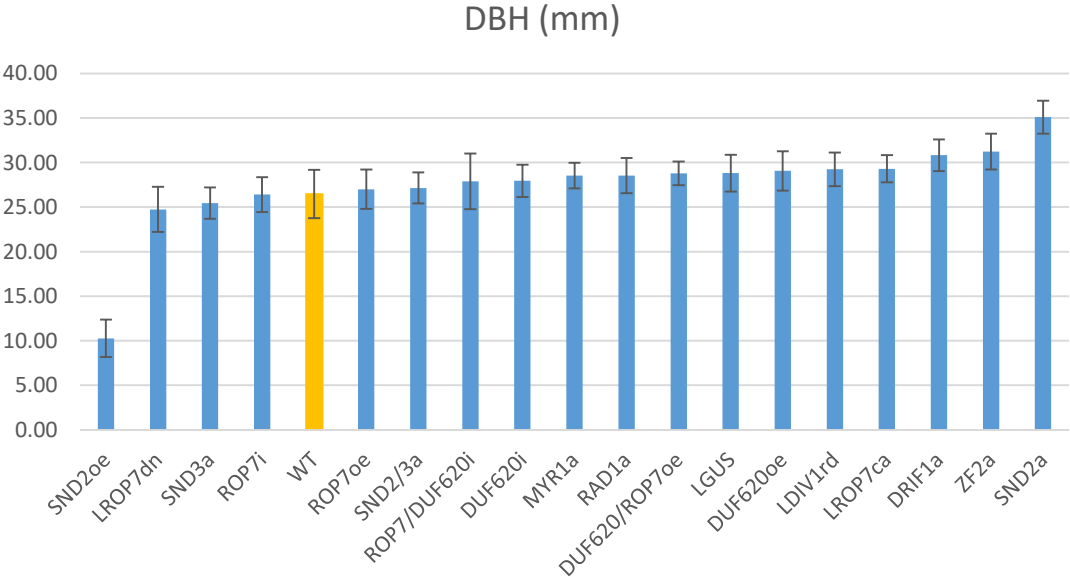
# Protein-Protein and Protein-DNA interactions linked to wood formation



Petzold et al. 2018

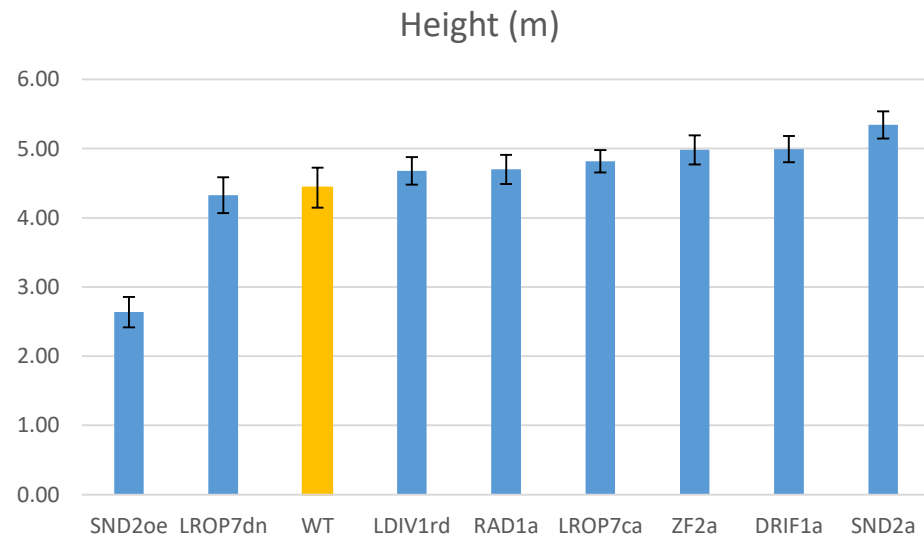
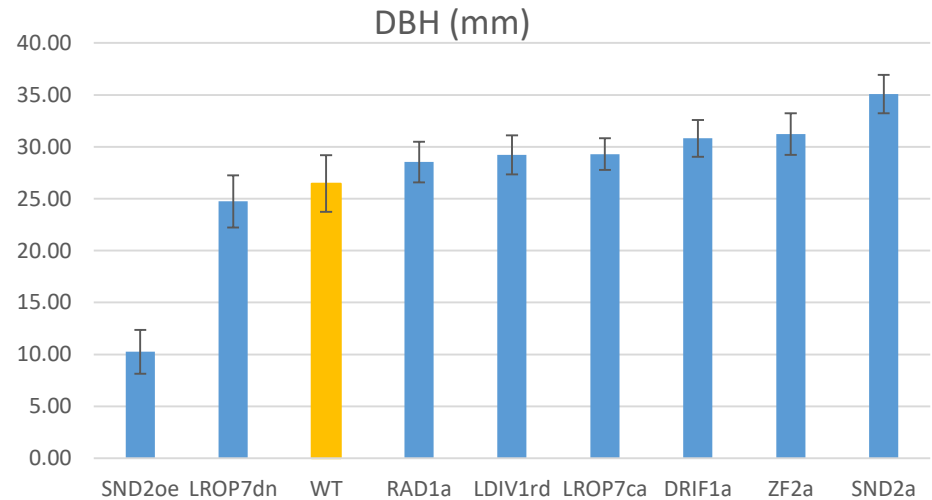
## Did we identify new regulators of wood formation?

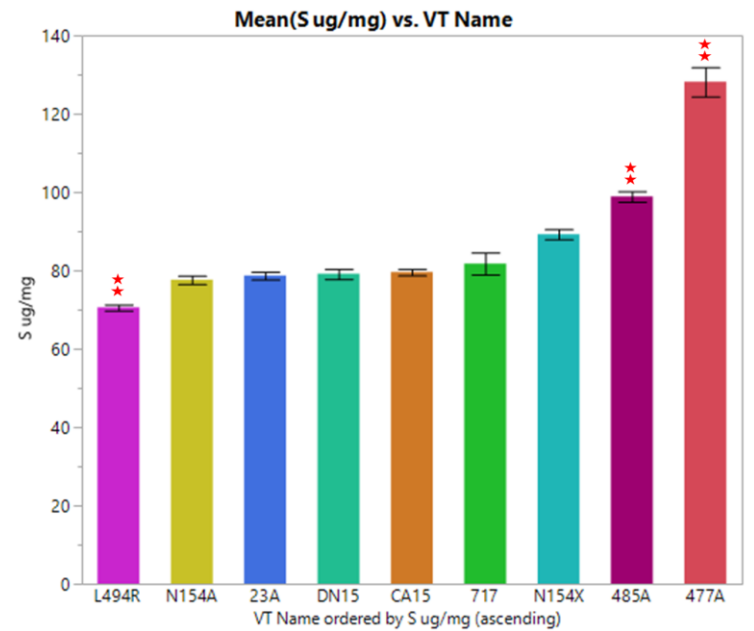
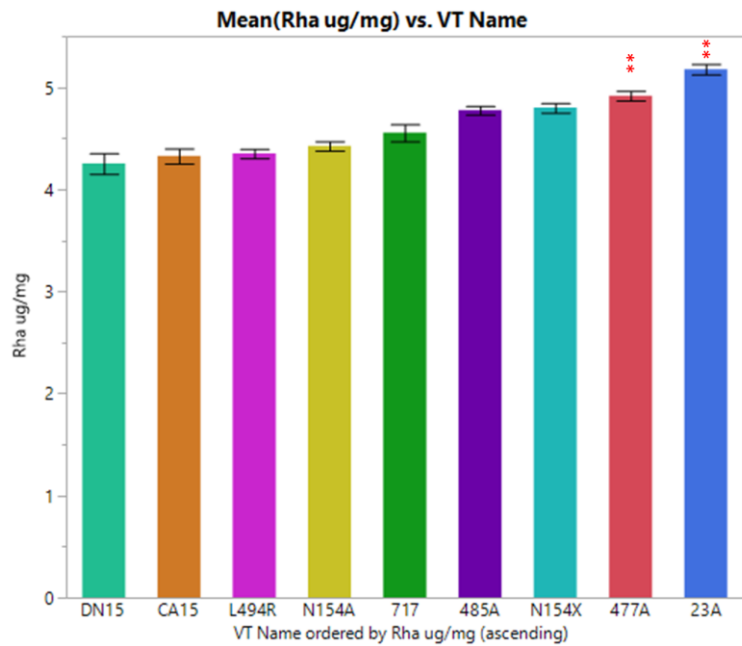
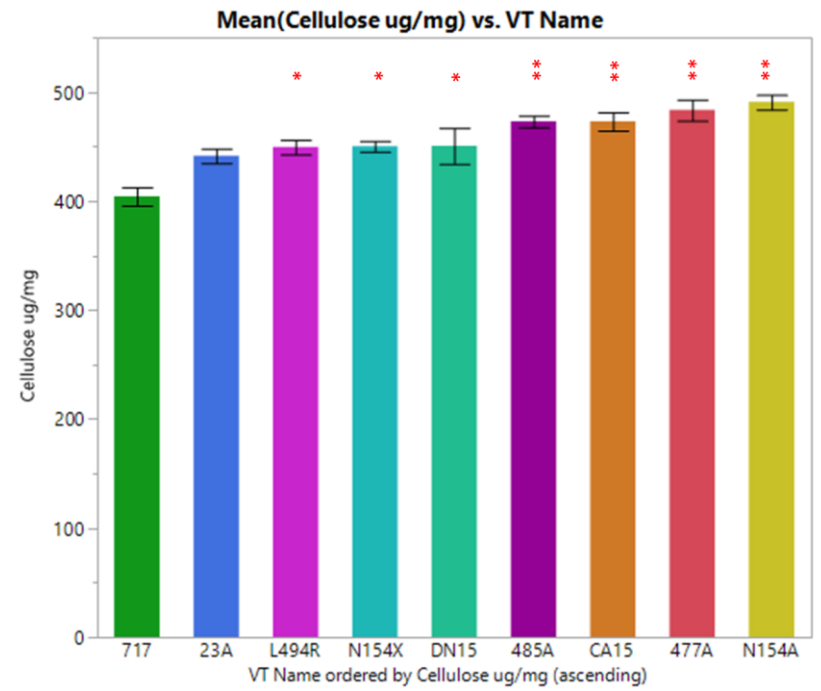
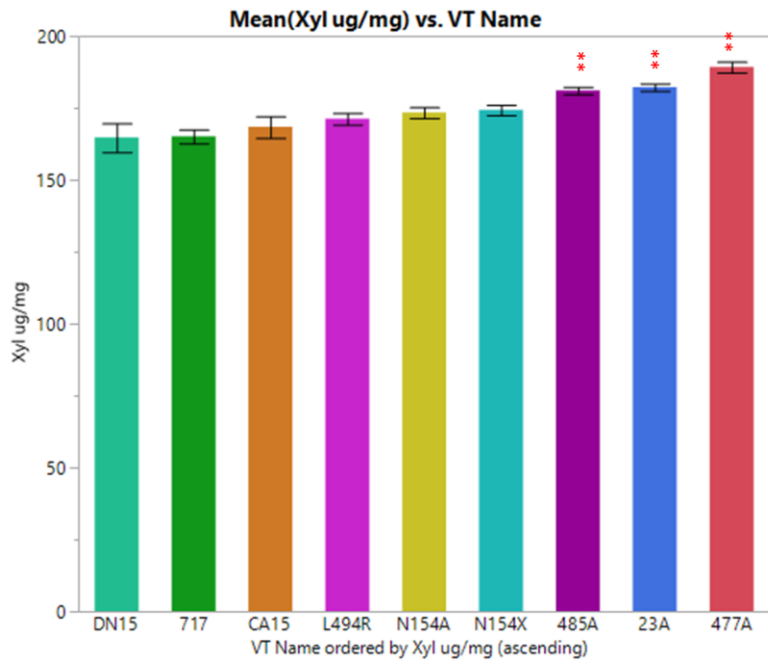
Transgenic field trial (Nov 2013-July 2017 ) included trees with 18 different constructs

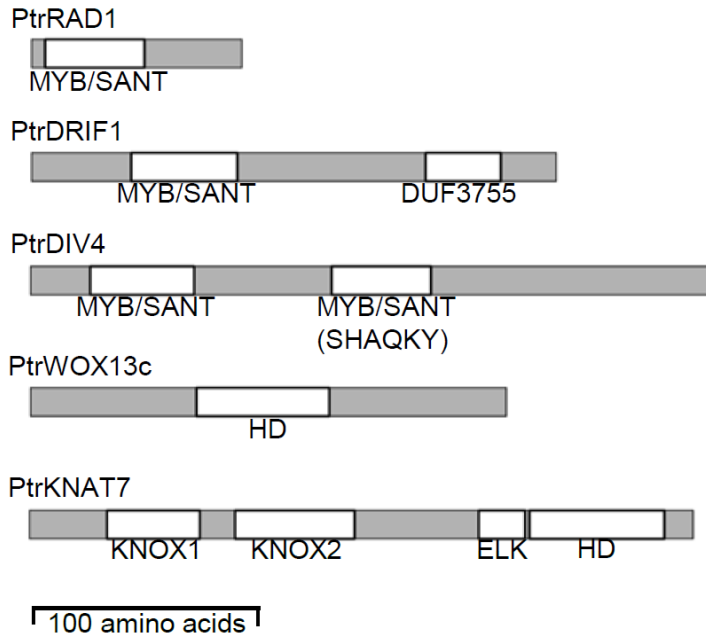


8 transgenic and WT were selected for wood chemistry analysis

- Performed by the GLBRC







	Y2H	Pulldown	BifC	Y2HB	BifCB	Localization
	+	+				V
	+		+			N
	+	+	+			N
	+	+	+			N
				+	+	N
	-	*				
				-		

Petzold et al. 2018

*LMX5::DIV1-SRDX*  
phenotypes



*LMX5::DIV1-SRDX*

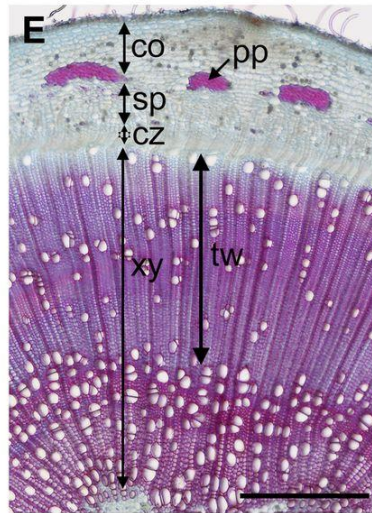
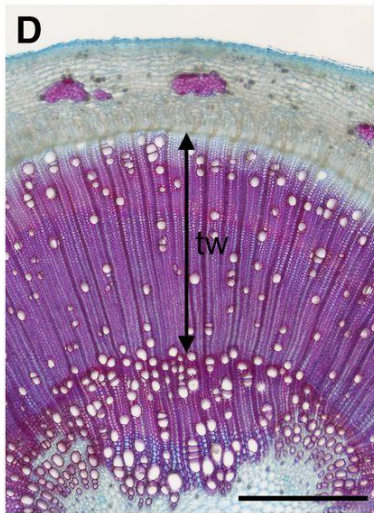
WT



miRNA-ARK2

OE-ARK2

OE-ARK2



Suzanne Gerttula et al. Plant Cell 2015;27:2800-2813



## Summary comments

- GRNs and integration with GWAS shows potential
  - As less biased approach to advance understanding of these complex processes in trees
  - For improving precision in identifying genes with key roles in phenology and growth
- SCW genes can have important roles in growth not related to wood formation
- Protein-Protein networks can identify novel regulators/regulatory complexes

- Rita Teixeira
- Hua Bai
- Xiaoyan Sheng
- Ayeshan Mahendra
- Steve Rigoulot
- Earl Petzold
- Bidisha Chanda
- Chengsong Zhao
- Xiaoyan Jia
- Mingzhe Zhao
- And many undergraduate students

- Jason Holliday
- Song Li
- Eric Beers
- Rich Helm
- **GLBRC**
- Cetin Yuceer
- Chuan-Yu Hsu

## Reynolds FRRC

- **Kyle Peer**
- **Deborah Bird**



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