### Supercalifragilistic Advances in Plant Genomics

### Jeremy Schmutz

HudsonAlpha Institute for Biotechnology Department of Energy Joint Genome Institute

> BERAC Science Talk October 25th, 2024



Huntsville, Alabama, USA



"Any sufficiently advanced technology is indistinguishable from magic."

Arthur C. Clarke 1973

Berkeley, California, USA

## In the beginning there was the human genome

In 1986 Department of Energy announced "Human Genome Initiative" in order to understand how radiation exposure affects DNA and is passed on in the germline

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## How did we sequence DNA?



ABI 377 slab gel





ABI 3730 capillary



1996

## Sequencing a whole genome (BAC vs WGS)?



- WGS requires:
  - As long as reads as possible
  - Positional information from pairs
  - Computational power
- WGS advantages:
  - Many fewer libraries
  - Streamlined production
  - Consistent & measurable quality

Research news | Published: 26 January 2001

### Weapons lab to develop Celera's new supercomputer

#### Robert Walgate

Genome Biology 2, Article number: spotlight-20010126-01 (2001) | Cite this article

733 Accesses | 1 Citations | 4 Altmetric | Metrics



LONDON Craig Venter, the CEO of <u>Celera Genomics</u> - which is on the verge of publishing the sequence of the human genome - has signed an agreement with <u>Sandia National Laboratory</u> in the US to develop the most powerful computer in the world within four years - and it'll be used for biology.

## What is assembly?

Shotgun ...ACCGTAAATGGGCTGATCATGCTTAAA sequence TGATCATGCTTAAACCCTGTGCATCCTACTG... Assembly ...ACCGTAAATGGGCTGATCATGCTTAAACCCTGTGCATCCTACTG...

Assembly = Computational reconstruction (aka model) of a linear representation of DNA from repeated sampling of sequence (reads) of variably quality & length



AAGCTAAGCCCAATTACGACCCAGATAGCTGGGGACGCGGCGTGATCGTAGCTGC

## Completing the human genome



Celebrating the human genome Craig Venter, Ari Patrinos, Francis Collins



46.9 MB DOE FINISHED CHROMOSOMES 323.1 Mb finished sequence 11.3% of human genome 41.0 MB



#### articles

## Initial sequencing and analysis of the 2001

International Human Genome Sequencing Consortium

\* A partial list of authors appears on the opposite page. Affiliations are listed at the end of the paper

The human genome holds an extraordinary trove of information about human development, physiology, medicine and evolution. Here we report the results of an international collaboration to produce and make freely available a draft sequence of the human genome. We also present an initial analysis of the data, describing some of the insights that can be gleaned from the sequence.

# Finishing the euchromatic sequence of the human genome

Marvin Frazier

2004

#### International Human Genome Sequencing Consortium

\* A list of authors and their affiliations appears in the Supplementary Information

Marvin Stodolsky

## What did JGI do with this sequencing capacity?

### **I FTTFRS**

#### The genome of Laccaria bicolor provides insights into mycorrhizal symbiosis

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### Genome sequence of the palaeopolyploid

Vol 452 6 March 2008 doi:10.1038/nature0655

#### soybean

Jeremy Schmutz<sup>1,2</sup>, Steven B, Cannon<sup>3</sup>, Jessica Schlueter<sup>4,5</sup>, Jianxin Ma<sup>5</sup>, Therese Mitros<sup>6</sup>, William Ne<sup>4</sup> David L. Hyten<sup>8</sup>, Qijian Song<sup>8,9</sup>, Jay J. Thelen<sup>10</sup>, Jianlin Cheng<sup>11</sup>, Dong Xu<sup>11</sup>, Uffe Hellsten<sup>2</sup>, Gregory D. RESEARCHARTICLES Yeisoo Yu<sup>13</sup>, Tetsuya Sakurai<sup>14</sup>, Taishi Umezawa<sup>14</sup>, Madan K. Bhattacharyya<sup>15</sup>, Devinder Sandhu<sup>16</sup> Babu Valliyodan<sup>17</sup>, Erika Lindquist<sup>2</sup>, Myron Peto<sup>3</sup>, David Grant<sup>3</sup>, Shengqiang Shu<sup>2</sup>, David Goodstein<sup>2</sup>, Ku

nature

Montona Futrell-Griggs<sup>5</sup>, Brian Abernathy<sup>5</sup>, Jianchang Du<sup>5</sup>, Zhixi Tian<sup>5</sup>, Liucun Zhu<sup>5</sup>, Navdeep Gill<sup>5</sup>, Tri Marc Libault<sup>17</sup>, Anand Sethuraman<sup>1</sup>, Xue-Cheng Zhang<sup>17</sup>, Kazuo Shinozaki<sup>14</sup>, Henry T. Nguyen<sup>17</sup>, Rod The Genome of Black Cottonwood. Perry Cregan<sup>8</sup>, James Specht<sup>18</sup>, Jane Grimwood<sup>1,2</sup>, Dan Rokhsar<sup>2</sup>, Gary Stacey<sup>10,17</sup>, Randy C. Shoemak

nature

# nature biotechnology

#### ARTICLES

Genome sequencing and analysis of the model grass Brachypodium distachyon

The International Brachypodium Initiative





nature

Science



nature

ARTICLE

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IFTTFRS

#### The Phaeodactylum genome reveals the evolutionary history of diatom genomes

Chris Bowler<sup>1,2</sup>, Andrew E. Allen<sup>1,3</sup>, Jonathan H. Badger<sup>3</sup>, Jane Grimwood<sup>4</sup>, Kamel Jabbari<sup>1</sup>, Alan Kuo<sup>5</sup>, Uma Maheswari<sup>1</sup>, Cindy Martens<sup>6</sup>, Florian Maumus<sup>1</sup>, Robert P. Otillar<sup>5</sup>, Edda Rayko<sup>1</sup>, Asaf Salamov<sup>3</sup>, Klaas Vandepoele<sup>6</sup>, Bank Beszteri<sup>7</sup>, Ansgar Gruber<sup>8</sup>, Marc Heijde<sup>1</sup>, Michael Katinka<sup>9</sup>, Thomas Mock<sup>10</sup> Klaus Valentin<sup>7</sup>, Fréderic Verret<sup>11</sup>, John A. Berges<sup>12</sup>, Colin Brownlee<sup>13</sup>, Jean-Paul Cadoret<sup>13</sup>, Anthony Chiovitti<sup>1</sup> Chang Jae Choi<sup>12</sup>, Sacha Coesel<sup>2</sup>†, Alessandra De Martino<sup>1</sup>, J. Chris Detter<sup>5</sup>, Colleen Durkin<sup>10</sup>, Angela Falciatore Jérome Fournet<sup>15</sup> Mixoshi Hanuta<sup>16</sup> Marie I. I. Huysman<sup>6,17</sup> Bethany D. Jenkins<sup>18</sup> Katerina Jiroutova Richard E. Jorgensen<sup>20</sup>, Yolaine Joubert<sup>15</sup>, Aaron Kaplan<sup>21</sup>, Nils Kröger<sup>22</sup>, Peter G. Kroth<sup>8</sup>, Julie La Roche<sup>1</sup> Erica Lindquist<sup>5</sup>, Markus Lommer<sup>23</sup>, Véronique Martin-Jézéquel<sup>15</sup>, Pascal J. Lopez<sup>1</sup>, Susan Lucas<sup>5</sup> Manuela Mangogna<sup>2</sup>, Karen McGinnis<sup>20</sup>, Linda K. Medlin<sup>7,11</sup>, Anton Montsant<sup>1,2</sup>, Marie-Pierre Oudot-Le Secq<sup>24</sup> Carolyn Napoli<sup>20</sup>, Miroslav Obornik<sup>19</sup>, Micaela Schnitzler Parker<sup>10</sup>, Jean-Louis Petit<sup>9</sup>, Betina M. Porcel<sup>9</sup>, Nicole Poulsen<sup>25</sup>, Matthew Robison<sup>16</sup>, Leszek Rychlewski<sup>26</sup>, Tatiana A. Rynearson<sup>27</sup>, Jeremy Schmutz Harris Shapiro<sup>5</sup>, Magali Siaut<sup>2</sup>†, Michele Stanley<sup>28</sup>, Michael R. Sussman<sup>16</sup>, Alison R. Taylor<sup>51,29</sup>, Assaf Vardi<sup>1,30</sup> Peter von Dassow<sup>31</sup>, Wim Vyverman<sup>17</sup>, Anusuya Willis<sup>14</sup>, Lucian S, Wyrwicz<sup>26</sup>, Daniel S, Rokhsar<sup>5</sup>, Jean Weissenbach<sup>9</sup>, E. Virginia Armbrust<sup>10</sup>, Beverley R. Green<sup>24</sup>, Yves Van de Peer<sup>6</sup> & Igor V. Grigoriev

> OPEN doi:10.1038/nature13308

### The genome of Eucalyptus grandis

Alexander A. Myburg<sup>1,2</sup>, Dario Grattapaglia<sup>3,4</sup>, Gerald A. Tuskan<sup>4,6</sup>, Ulfe Hellsten<sup>4</sup>, Richard D. Hayes<sup>5</sup>, Jane Grimwood<sup>7</sup>, Jerry Jenkins<sup>7</sup>, Erika Lindquist<sup>2</sup>, Hope Tice<sup>2</sup>, Diane Bauer<sup>2</sup>, David M. Goodstein<sup>3</sup>, Inna Dubchak<sup>2</sup>, Alexandre Dollakov<sup>3</sup>, Eshchar Mizrachi<sup>1,2</sup>, Anand R. K. Klullan<sup>1,2</sup>, Steven G. Hussey<sup>1,2</sup>, Desre Pinard<sup>1,2</sup>, Karen van der Merwe<sup>1,2</sup>, Pooja Singh<sup>1,2</sup>, Ida van Jaarsveld<sup>4</sup>, Orzenil B. S. Kluu-Juniof<sup>2</sup>, Roberto G. Togawa<sup>3</sup>, Mariila R. Papas<sup>3</sup>, Danielle A. Frait<sup>3</sup>, Carolina P. Sansalon<sup>1</sup> Cesar D. Petroli<sup>3</sup>, Xiaohan Yang<sup>6</sup>, Priya Ranjan<sup>6</sup>, Timothy J. Tschaplinski<sup>6</sup>, Chu-Yu Ye<sup>6</sup>, Ting Li<sup>6</sup>, Lieven Sterck<sup>10</sup>, Kevin Vanneste<sup>10</sup> Cesar D', retroit "Auonan Tang", r'hya Rangan", innotiny i senapiniski "Cut- 'u te', ing U, Leven Setrec, "Aewi Nameset" Forent Mural", Marcia Soler", Héléne San Chemente", Najib Said", 'Hua Cassan "Wang", Christopher Danand", Charles A. Hefer<sup>3,1</sup>, Brich Bornberg-Bauer", Anna R. Kersting<sup>4,45</sup>, Kelly Wining<sup>16</sup>, Vindhya Amarasinghe<sup>10</sup>, Marin Ranik<sup>16</sup>, Sushma Naithan<sup>17,8</sup>, Justin Bert", 'Alexander E. Boyd<sup>18</sup>, Aron Liston<sup>12,8</sup>, Joseph W, Spatzlor<sup>21,87</sup>, Palitha Dharmwardhana<sup>17</sup>, Rajani Raja<sup>17</sup>, Christopher Sullivan<sup>18</sup>, Elison Romanel<sup>19,50,27</sup>, Marcio Alves - Ferreira<sup>17</sup>, Carsten Külheim<sup>27</sup>, Willam Foley<sup>72</sup>, Vietor Carocha<sup>12,73,41</sup>, togge Hurdin<sup>23,41</sup>, David Kurlar<sup>25,50</sup>, Marcio Alves - Ferreira<sup>17</sup>, Carsten Külheim<sup>27</sup>, Willam Foley<sup>72</sup>, Vietor Carocha<sup>12,73,41</sup>, togge Hurdin<sup>23,41</sup>, David Kurlar<sup>25,50</sup>, Marcio Alves - Ferreira<sup>17</sup>, Carsten Külheim<sup>27</sup>, Willam Foley<sup>72</sup>, Philippe Rigault<sup>70</sup>, Josquin Tibbits<sup>10</sup>, Antanas Spokevicius<sup>18</sup>, Rebeca C, Jones<sup>27</sup>, Dorothy A, Steane<sup>17,25</sup>, René E, Vallancourt<sup>17</sup>, Kullan Kurlar<sup>15,150</sup>, Kene Kurlar<sup>15,150</sup> Brad M. Potts<sup>32</sup>, Fourie Joubert<sup>3,8</sup>, Kerrie Barry<sup>5</sup>, Georgios J. Pappas Jr<sup>34</sup>, Steven H. Strauss<sup>16</sup>, Pankaj Jaiswal<sup>1</sup> Jacqueline Grima-Pettenati<sup>12</sup>, Jérôme Salse<sup>11</sup>, Yves Van de Peer<sup>2,10</sup>, Daniel S. Rokhsar<sup>5</sup> & Jeremy Schmutz<sup>5,7</sup>

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ARTICIES

### The Sorghum bicolor genome and the

diversification of grasses Andrew H. Paterson<sup>1</sup>, John E. Bowers<sup>1</sup>, Rémy Bruggmann<sup>2</sup>, Inna Dubchak<sup>3</sup>, Jane Grimwood<sup>4</sup>, Heidrun Gundlach<sup>5</sup>, Georg Haberer<sup>5</sup>, Uffe Hellsten<sup>3</sup>, Therese Mitros<sup>6</sup>, Alexander Poliakov<sup>3</sup>, Jeremy Schmutz<sup>4</sup>, Manuel Spannagl<sup>5</sup>, Haibao Tang<sup>1</sup>, Xiyin Wang<sup>1,7</sup>, Thomas Wicker<sup>8</sup>, Arvind K. Bharti<sup>2</sup>, Jarrod Chapman<sup>3</sup>, F. Alex Feltus<sup>1,9</sup>, Udo Gowik<sup>10</sup> Igor V. Grigoriev<sup>5</sup>, Eric Lyons<sup>11</sup>, Christopher A, Maher<sup>12</sup>, Mihaela Martis<sup>5</sup>, Apurya Narechania<sup>12</sup>, Robert P, Otillar<sup>1</sup> Bryan W. Penning<sup>13</sup>, Asaf A. Salamov<sup>3</sup>, Yu Wang<sup>5</sup>, Lifang Zhang<sup>12</sup>, Nicholas C. Carpita<sup>14</sup>, Michael Freeling<sup>11</sup>, Alan R. Gingle<sup>1</sup>, C. Thomas Hash<sup>15</sup>, Beat Keller<sup>8</sup>, Patricia Klein<sup>16</sup>, Stephen Kresovich<sup>17</sup>, Maureen C. McCann<sup>1</sup> Ray Ming<sup>18</sup>, Daniel G. Peterson<sup>1,19</sup>, Mehboob-ur-Rahman<sup>1,20</sup>, Doreen Ware<sup>12,21</sup>, Peter Westhoff<sup>10</sup> Klaus F. X. Mayer<sup>5</sup>, Joachim Messing<sup>2</sup> & Daniel S. Rokhsar<sup>3,4</sup>

Sequencing of diverse mandarin, pummelo and orange genomes reveals complex history of admixture during citrus domestication

G Albert Wu<sup>1,29</sup>, Simon Prochnik<sup>1,29</sup>, Jerry Jenkins<sup>2</sup>, Jerome Salse<sup>3</sup>, Uffe Hellsten<sup>1</sup>, Florent Murat Xavier Perrier<sup>4</sup>, Manuel Ruiz<sup>4</sup>, Simone Scalabrin<sup>5</sup>, Javier Terol<sup>6</sup>, Marco Aurélio Takita<sup>7</sup>, Karine Labadie<sup>8</sup>, Julie Poulain<sup>8</sup>, Arnaud Couloux<sup>8</sup>, Kamel Jabbari<sup>8</sup>, Federica Cattonaro<sup>5</sup>, Cristian Del Fabbro<sup>5</sup>, Sara Pinosio<sup>5</sup> Andrea Zuccolo<sup>5,9</sup>, Jarrod Chapman<sup>1</sup>, Jane Grimwood<sup>2</sup>, Francisco R Tadeo<sup>6</sup>, Leandro H Estornell<sup>6</sup>, Juan V Muñoz-Sanz<sup>6</sup>, Victoria Ibanez<sup>6</sup>, Amparo Herrero-Ortega<sup>6</sup>, Pablo Aleza<sup>10</sup>, Julián Pérez-Pérez<sup>11,12</sup> Daniel Ramón<sup>11</sup>, Dominique Brunel<sup>8,13</sup>, François Luro<sup>14</sup>, Chanxian Chen<sup>15,28</sup>, William G Farmerie<sup>16</sup>, Brian Desany<sup>17</sup>, Chinnappa Kodira<sup>17</sup>, Mohammed Mohiuddin<sup>17</sup>, Tim Harkins<sup>17,28</sup>, Karin Fredrikson<sup>11</sup> Paul Burra<sup>15,19</sup>, Alexandre Lomsadze<sup>18,19</sup>, Mark Borodovsky<sup>18-20</sup>, Giuseppe Reforgiato<sup>21</sup>, Juliana Freitas-Astúa<sup>7,22</sup> Francis Quetier<sup>8,23</sup>, Luis Navarro<sup>19</sup>, Mikeal Roose<sup>24</sup>, Patrick Wincker<sup>8,23,25</sup>, Jeremy Schmutz<sup>2</sup>, Michele Morgante<sup>5,26</sup>, Marcos Antonio Machado<sup>7</sup>, Manuel Talon<sup>6</sup>, Olivier Jaillon<sup>8,23,25</sup>, Patrick Ollitrault<sup>4</sup>, Frederick Gmitter<sup>15</sup> & Daniel Rokhsar<sup>1,27</sup>



## G. A. Tuskan,<sup>1,2\*</sup> S. DiFazio,<sup>1,4</sup> † S. Jansson,<sup>5</sup> † J. Bohlmann,<sup>4</sup> † I. Grigoriev,<sup>9</sup> † U. Hellsten,<sup>9</sup> † N. Putnam,<sup>8</sup> † S. Ralph,<sup>4</sup> † S. Rombauts,<sup>50</sup> † A. Salamov,<sup>4</sup> † J. Schein,<sup>13</sup> † L. Sterck,<sup>15</sup> † A. Aerts,<sup>1</sup> R. B. Bhaleroo,<sup>5</sup> R. P. Bhaleroo,<sup>12</sup> D. Blaudez,<sup>13</sup> W. Boerian,<sup>15</sup> A. Brunu<sup>13</sup> A. Brunner,<sup>14</sup>

Populus trichocarpa (Torr. & Grav)

R. R. Bhalerao,\* R. P. Bhalerao,\* D. Blaudez,\*\* W. Boorjan,\*\* A. Brun,\*\* A. Brunner,\*\* W. Buroy,\* M. Campbell,\*\* J. Cattorine,\*\* D. Blaudez,\*\* W. Boorjan,\*\* A. Brunner,\*\* P. M. Coutinho,\*\* J. Couttiner,\*\* S. Covert,\*\* O. Cronk,\* R. Conningham,\*\* El Davis,\*\* S. Degroser,\*\* A. Dajscinfi,\*\* C. Catharphilis,\*\* J. Deters\*\* D. Bioks,\*\* J. Bubchak,\*\* S. Duplessi,\*\* J. Exhting,\*\* B. Ellis,\* K. Gendler,\*\* D. Goodstein,\*\* M. Girbakov,\*\* J. Grimmood,\*\* A. Grover,\*\* L. Gunter,\* B. Anaberger, R. Heinz,\*\* V. Healtrita, 1,\*\*\* B. Henriss,\*\* D. Holligan,\*\* R. Holt,\*\* W. Huang, \*\* Listan-Faridi,\*\* S. Jones,\*\* M. B. Ineors-Rhoades,\*\* R. Jongensen,\*\* C. Dohk\*\*, S. Kantonian,\*\* C. Napadi,\*\* D. Hearting, \*\* Landes,\*\* M. Kirk,\*\* A. Kohlex,\*\* U. Kalluri,\* F. Larimer,\* J. Leebens-Mack,\*\* J.-C. Leple,\*\* P. Locascio,\* Y. Lov,\* S. Lucz,\*\* R. Martin,\*\* B. Anoniani,\*\* C. Napadi,\*\* D. R. Kirkaari,\*\* C. Neston,\*\* K. Hiemmen,\*\* O. Hilsson,\*\* V. Pereda,\*\* G. Peter,\*\* K. Philippe,\* G. Flatta,\*\* A. Kohlex,\*\* B. Kazumovskaya,\* R. Richardion,\*\* C. Knalad,\*\* S. K. Hull,\*\* S. Koses,\*\* G. Nabav,\*\* J. Schmatz,\*\* J. Schrader,\*\* B. Segerman,\*\* H. Shin,\*\* A. Siddiqui,\*\* F. Steky,\*\* A. Ferry,\* C. J. Tau,\*\* C. Buethacher,\*\* R. J. Manehamer,\*\* J. Leabers,\*\* Kull,\*\* S. Koses,\*\* L. Schmatz,\*\* B. Schrader,\*\* B. Segerman,\*\* H. Shin,\*\* A. Siddiqui,\*\* F. Steky,\*\* A. Ferry,\* C. J. Tau,\*\* C. Washacher,\*\* B. Segerman,\*\* H. Shin,\*\* A. Siddiqui,\*\* F. Steky,\*\* A. Ferry,\*\* C. J. Tau,\*\* C. Washacher,\*\* B. Segerman,\*\* G. Sandberg,\*\* Y. Van de Peer,\*\*\* D. Rokhar\*\*\* B. Segerma,\*\* B. Steky,\*\* K. Hull,\*\*, S. Seser,\*\* G. Sang,\*\*\* T. Yin,\* C. Douglas,\* M. Marra,\*\* G. Sandberg,\*\*\* Y. Van de Peer,\*\*\* D. Rokhar\*\*\*\*

### LETTER

#### Repeated polyploidization of Gossypium genomes and the evolution of spinnable cotton fibres

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## Post-Sanger genome sequencing











Solexa GA 2006

Illumina 2500 2012

PacbBio RS 2011

PacbBio RSII 2013

PacbBio Sequel 2016

Short read platforms

- + super inexpensive data
- + low error rate (99.5% accuracy)
- + metric tons of it
- 76 to 150 bp reads
- Short fragments (400bp)

Long read platforms

- expensive data (its all relative)
- high error rate (82% accuracy)
- low production
- + 2,000 50,000 bp reads
- + Greatly reduce assembly complexity

## New science with long reads

### genetics

ARTICLES https://doi.org/10.1038/s41588-019-0405-z

OPEN

### The genome sequence of segmental allotetraploid peanut *Arachis hypogaea*

David J. Bertioli (12.3.30\*, Jerry Jenkins) 4.30, Josh Clevenger 1.2.3.30, Olga Dudchenko 5, Dongying Gao1, Guillermo Seijo<sup>6,7</sup>, Soraya C. M. Leal-Bertioli<sup>12.8</sup>, Longhui Ren<sup>9</sup>, Andrew D. Farmer<sup>10</sup>, Manish K. Pandey 1, Sergio S. Samoluk<sup>6,7</sup>, Brian Abernathy<sup>1</sup>, Gaurav Agarwal<sup>8</sup>, Carolina Ballén-Taborda<sup>2</sup>, Connor Cameron<sup>10</sup>, Jacqueline Campbell <sup>10</sup>, Carolina Chavarro<sup>12</sup>, Annapurna Chitikineni<sup>11</sup>, Ye Chu<sup>3</sup>, Sudhansu Dash<sup>10</sup>, Moaine El Baidouri<sup>14,51</sup>, Baozhu Guo<sup>6</sup>, Wei Huang<sup>12</sup>, Kyung Do Kim<sup>11</sup>, Walid Korani <sup>10</sup>, Sophie Lanciano<sup>15,18,19</sup>, Christopher G. Lui<sup>5</sup>, Marie Mirouze <sup>15,18,19</sup>, Márcio C. Moretzsohn<sup>20</sup>, Melanie Pham<sup>5</sup>, Jin Hee Shin<sup>117</sup>, Kenta Shirasawa <sup>21</sup>, Senjuti Sinharoy<sup>22</sup>, Avinash Sreedasyam <sup>4</sup>, Nathan T. Weeks <sup>23</sup>, Xinyou Zhang<sup>24,25</sup>, Zheng Zheng<sup>24,25</sup>, Zigi Sun<sup>24,25</sup>, Lutz Froenicke<sup>26</sup>, Erez L. Aiden<sup>5</sup>, Richard Michelmore<sup>26</sup>, Rajeev K. Varshney <sup>11</sup>, C. Corley Holbrok<sup>27</sup>, Ethalinda K. S. Cannon <sup>12</sup>, Brian E. Scheffler <sup>28</sup>, Jane Grimwood<sup>4</sup>, Peggy Ozias-Akins<sup>2,13</sup>, Steven B. Cannon <sup>62,3,31</sup>, Scott A. Jackson <sup>61,2,3,31</sup>\* and Jeremy Schmutz <sup>64,29,31\*</sup>

Inbred Tetraploid (2x)



Outbred Diploid (2x) Partial resolved 2nd nature

#### ARTICLE

https://doi.org/10.1038/s41467-021-24328-w OPEN

Four chromosome scale genomes and a pangenome annotation to accelerate pecan tree breeding

John T. Lovello <sup>1,1584</sup>, Nolan B. Bentley <sup>2,15</sup>, Gaurab Bhattarai<sup>3,15</sup>, Jerry W. Jenkins <sup>1,15</sup>, Avinash Sreedasyam <sup>1,15</sup>, Yanina Alarcon <sup>4</sup>, Clive Bock<sup>5</sup>, Lori Beth Boston<sup>1</sup>, Joseph Carlson<sup>6</sup>, Kimberly Cervantes<sup>7</sup>, Kristen Clermont<sup>8</sup>, Sara Duke<sup>9</sup>, Nick Krom<sup>4</sup>, Keith Kubenka<sup>10</sup>, Sujan Mamidi<sup>1</sup>, Christopher P. Mattison <sup>8</sup>, Maria J. Monteros <sup>4</sup>, Cristina Pisani<sup>5</sup>, Christopher Plott<sup>1</sup>, Shanmugam Rajasekar<sup>11</sup>, Hormat Shadgou Rhein<sup>7</sup>, Charles Rohla<sup>4</sup>, Mingzhou Song<sup>12</sup>, Rolston St. Hilaire<sup>13</sup>, Shengqiang Shuo <sup>6</sup>, Lenny Wells<sup>14</sup>, Jenell Webber<sup>1</sup>, Richard J. Heerema <sup>12</sup>, Patricia E. Klein <sup>9</sup>, Patrick Conner<sup>14</sup>, Xinwang Wang<sup>10</sup>, L. J. Grauke <sup>10</sup>, Jane Grimwood <sup>1</sup>, Jeremy Schmutz <sup>1688</sup> & Jennifer J. Randall<sup>788</sup>

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### Article Genomic mechanisms of climate adaptation in polyploid bioenergy switchgrass

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Check for updates
Open access
Published online: 27 January 2
Accepted: 16 December 2020

### nature

John T. Lovell<sup>1345</sup>, Alice H. MacQueen<sup>2,38</sup>, Sujan Mamidi<sup>138</sup>, Jason Bonnette<sup>2,38</sup>, Jerry Jenkins<sup>138</sup>, Joseph D. Napier<sup>2</sup>, Avinash Sreedasyam<sup>3</sup>, Adam Healey<sup>1</sup>, Adam Session<sup>3,4</sup>, Shengqiang Shu<sup>2</sup>, Kerie Barry<sup>3</sup>, Stacy Bonos<sup>6</sup>, LoriBeth Boston<sup>1</sup>, Christopher Daum<sup>3</sup>, Shweta Deshande<sup>3</sup>, Aren Ewing<sup>3</sup>, Paul P. Grabowski<sup>1</sup>, Taslima Haque<sup>6</sup>, Melanie Harrison<sup>4</sup>, Jiming Jiang<sup>9</sup>, Dave Kudma<sup>4</sup>, Anna Lipzen<sup>9</sup>, Thomas H. Pendergast IV<sup>610,17</sup>, Chris Plott<sup>1</sup>, Peng Qi<sup>9</sup>, Christopher A. Saski<sup>10</sup>, Eugene V. Shakirov<sup>2,13</sup>, David Sims<sup>1</sup>, Manoj Sharma<sup>4</sup>, Rita Sharma<sup>3</sup>, Ada Stewart<sup>1</sup>, Vasanth R. Singan<sup>3</sup>, Yuhong Tang<sup>16</sup>, Sandra Thibivillier<sup>7</sup>, Jenell Webber<sup>1</sup>, Xiaoyu Weng<sup>4</sup>, Melissa Williams<sup>1</sup>, Guohong Albert Wu<sup>3</sup>, Yuko Yoshinaga<sup>3</sup>, Matthew Zane<sup>9</sup>, Li Zhang<sup>4</sup>, Jiyi Zhang<sup>16</sup>, Kathrine D. Behrman<sup>1</sup>, Arvid R. Boe<sup>18</sup>, Philip A. Fay<sup>30</sup>, Rois R. Fritsch<sup>27,1</sup>, Julio Lastrow<sup>21</sup>, John Lioyd-Reilley<sup>27</sup>, Juan Manuel Martinez. Reyna<sup>23</sup>, Roser Matamala<sup>21</sup>, Robert B. Mitchell<sup>24</sup>, Francis M. Rouquette Jr<sup>28</sup>, Pamela Ronald<sup>28,27</sup>, Malay Saha<sup>30</sup>, Christian M. Tobia<sup>35</sup>, Michael Udvard<sup>110</sup>, Rod A. Wing<sup>4</sup>, Yanqi Wu<sup>27</sup>, Laura E. Bartley<sup>20,21</sup>, Michael Casler<sup>22,23</sup>, Katrien M. Devo<sup>30,101,43</sup>, Javid B. Lowry<sup>23,5</sup>, Daniel S. Rokhsar<sup>2,43,53,7</sup>, Jane Grimwood<sup>4</sup>, Thomas E. Juenger<sup>22,58</sup> & Jeremy Schmutz<sup>13,58</sup>



Outbred Tetraploid (4x) Blended single haplotype

### genetics

ARTICLES https://doi.org/10.1038/s41588-020-0614-5

### Check for upda

#### Genomic diversifications of five *Gossypium* allopolyploid species and their impact on cotton improvement

Z. Jeffrey Chen <sup>12,14</sup><sup>SD</sup>, Avinash Sreedasyam <sup>3,34</sup>, Atsumi Ando<sup>1,34</sup>, Qingxin Song<sup>12,14</sup>, Luis M. De Santiago <sup>4,14</sup>, Amanda M. Hulse-Kemp<sup>5</sup>, Mingquan Ding<sup>15</sup>, Wenxue Ye<sup>2</sup>, Ryan C. Kirkbride<sup>3</sup>, Jerry Jenkins<sup>3</sup>, Christopher Plott<sup>3</sup>, John Lovell<sup>3</sup>, Yu-Ming Lin<sup>4</sup>, Robert Vaughn<sup>4</sup>, Bo Liu<sup>4</sup>, Sheron Simpson<sup>7</sup>, Brian E. Scheffler <sup>5</sup>, Li Wen<sup>8</sup>, Christopher A. Saski<sup>8</sup>, Corrinne E. Grover <sup>5</sup>, Guanjing Hu<sup>9</sup>, Justin L. Conover <sup>9</sup>, Joseph W. Carlson<sup>10</sup>, Shengqiang Shu<sup>9,10</sup>, Lori B. Boston<sup>3</sup>, Melissa Williams<sup>3</sup>, Daniel G. Peterson<sup>11</sup>, Keith McGee<sup>12</sup>, Don C. Jones<sup>13</sup>, Jonathan F. Wendel <sup>9</sup>, David M. Stelly<sup>64</sup>, Jane Grimwood<sup>5,128</sup> and Jeremy Schmutz<sup>310</sup>

### Inbred Tetraploid (2x)





- Enabled by longer primary reads (up to 200kb)
- Better data collection on new instruments
- New analysis techniques

 2023 – new instruments that collect 13x the data for 1/3 cost

PacBio Revio 2023

## DNN model for consensus

### DeepConsensus: Gap-Aware Sequence Transformers for Sequence Correction

Gunjan Baid<sup>1\*</sup>, Daniel E. Cook<sup>1\*</sup>, Kishwar Shafin<sup>1</sup>, Taedong Yun<sup>1</sup>, Felipe Llinares-López<sup>1</sup>, Quentin Berthet<sup>1</sup>, Aaron M. Wenger<sup>2</sup>, William J. Rowell<sup>2</sup>, Maria Nattestad<sup>1</sup>, Howard Yang<sup>1</sup>, Alexey Kolesnikov<sup>1</sup>, Armin Töpfer<sup>2</sup>, Waleed Ammar<sup>1</sup>, Jean-Philippe Vert<sup>1</sup>, Ashish Vaswani<sup>1</sup>, Cory Y. McLean<sup>1</sup>, Pi-Chuan Chang<sup>1</sup>^, Andrew Carroll<sup>1</sup>^







**Pi-Chang Chang** 

Andrew Carroll

Google Al





Nature Biotechnology 41, 232–238 (2023)

## Quality of reads



### Science

Info & Affiliations

HOME > SCIENCE > VOL. 376. NO. 6588 > FROM TELOMERE TO TELOMERE:

genetic state of human repeat elements

SPECIAL ISSUE RESEARCH ARTICLE HUMAN GENOMICS

From telomere to telomere: The transcriptional and epi-

SAVANNAH J. HOYT 🍈 , JESSICA M. STORER 🍈 , GABRIELLE A. HARTLEY, PATRICK G. S. GRADY 🍈 , ARIEL GERSHMAN 🍈 , LEONARDO G. DE LIMA 🝈 , CHARLES LIMOUSE REZA HALABIAN 🍈 , LUKE WOJENSKI 🍈 , MATIAS RODRIGUEZ 🍈 , NICOLAS ALTEMOSE 🍈 , ARANG RHIE 🍈 , LEIGHTON J. CORE 🍈 , JENNIFER L. GERTON 🍈 , WOJ-

CIECH MAKALOWSKI (D), DANIEL OLSON (D), JEB ROSEN (D), ARIAN F. A. SMIT (D), AARON F. STRAIGHT (D), MITCHELL R. VOLLGER (D), TRAVIS J. WHEELER (D)

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linked to diseases.

The New Hork Eimes

Home / 2022 / May / Karen Mica Named one of TIME's 100 Most Influential People of 2022 Karen Miga Named one of TIME's 100 Most Influential People of 2022 May 23, 2022

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#### Karen Miga, assistant professor of biomolecula engineering at UC Santa Cruz, was named one of 100 most influential people of 2022 by TIM nor was announced on May 23.

figa and her colleagues, Adam Phillippy, Eva Eichler, and Michael Schatz, led an international team of scientists - the Telomere-to-Telomere (T2T) Consortium - to complete the first gapless sequence of the human genome. Parts of the human genome are now available to stud for the first time, allowing researchers to better inderstand genetic diseases, human diversity and evolution The gaps now filled by the new sequence in



### Scientists Have Finally Sequenced the Complete Human Genome – And Revealed New Genetic Secrets

TOPICS DNA Genetics Genome Popular IIC Berkeley

are probably functional, and many new variants that may be



Find it on github: jtlovell/GENESPACE

Three lines of R to compare multiple complex genomes

MICHAEL C. SCHATZ 🍈 , EVAN E. EICHLER 🍈 , ADAM M. PHILLIPPY 🍈 , WINSTON TIMP 🍈 , KAREN H. MIGA 🍈 , AND RACHEL J. O'NEILL 📵 Authors fewer

Lovell & Grimwood, Nature News and Views, 2022

### 99+% of this is from HIFI alone

Scientists Finish the Human Genome

By Public Affairs



The complete genome uncovered more than 100 new genes that

the entire short arms of five human chromosomes and cover some of the most complex regions of the genome. These include highly repetitive DNA sequences found in ane



### COMPLEXITY

## Inbred genome references of small plants

Year	Build	Assembled Size	Contig Number	Contig N50 (MB)
2007	V1 WGS Sanger <sup>1</sup>	688 MB	12,873	0.2
2013	Rio AllPaths Illumina	404 MB	49,137	0.02
2015	V3 WGS Improved <sup>2</sup>	675 MB	4,783	1.3
2016	Rio RSII <sup>3</sup>	729 MB	3,830	0.4
2018	V4 PacBio Sequel <sup>4</sup>	721 MB	411	17.7
2020	V5 PacBio Sequel2 <sup>4</sup>	720 MB	89	50.7
2022	V6 PacBio Hi-Fi <sup>4</sup>	718 MB	21	55.0



<sup>1</sup> Paterson, Nature 2009
<sup>2</sup> McCormick, Plant 2017
<sup>3</sup> Cooper, BMC genomics 2019
<sup>4</sup> Unpublished

## Poplar experiments

#### RESEARCH ARTICLE

#### The Genome of Black Cottonwood, Populus trichocarpa (Torr. & Gray)

G. A. Tuskan<sup>1,3,\*</sup>, S. DiFazio<sup>1,4,†</sup>, S. Jansson<sup>5,†</sup>, J. Bohlmann<sup>6,†</sup>, I. Grigoriev<sup>9,†</sup>, U. Hellsten<sup>9,†</sup>, N. Putnam<sup>9,†</sup>, S. Ralph<sup>6,†</sup>, S. Rombauts<sup>10,†</sup>, A. Salamov<sup>9,†</sup>, J. Schein<sup>11,†</sup>, L. Sterck<sup>10,†</sup>, A. Aerts<sup>9</sup>, R. R. Bhalerao<sup>5</sup>, R. P. Bhalerao<sup>12</sup>, D. Blaudez<sup>13</sup>, W. Boerjan<sup>10</sup>, A. Brun<sup>13</sup>, A. Brunner<sup>14</sup>, V. Busov<sup>15</sup>, M. Campbell<sup>16</sup>, J. Carlson<sup>17</sup>, M Chalot<sup>13</sup>, J. Chapman<sup>9</sup>, G.-L. Chen<sup>2</sup>, D. Cooper<sup>6</sup>, P. M. Coutinho<sup>19</sup>, J. Couturier<sup>13</sup>, S. Covert<sup>20</sup>, Q. Cronk<sup>7</sup>, R. Cunningham<sup>1</sup>, J. Davis<sup>22</sup>, S. Degroeve<sup>10</sup>, A. Déjardin<sup>23</sup>, C. dePamphilis<sup>18</sup>, J. Detter<sup>9</sup>, B. Dirks<sup>24</sup>, I. Dubchak<sup>9,25</sup>, S. Duplessis<sup>13</sup>, J. Ehlting<sup>7</sup>, B. Ellis<sup>6</sup>, K. Gendler<sup>26</sup>, D. Goodstein<sup>9</sup>, M. Gribskov<sup>27</sup>, J. Grimwood<sup>28</sup>, A. Groover<sup>29</sup>, L. Gunter<sup>1</sup>, B. Hamberger<sup>7</sup>, B. Heinze<sup>30</sup>, Y. Helariutta<sup>12,31,33</sup>, B. Henrissat<sup>19</sup>, D. Holligan<sup>21</sup>, R. Holt<sup>11</sup>, W. Huang<sup>9</sup>, N. Islam-Faridi<sup>34</sup>, S. Jones<sup>11</sup>, M. Jones-Rhoades<sup>35</sup>, R. Jorgensen<sup>26</sup>, C. Joshi<sup>15</sup>, J. Kangasjärvi<sup>32</sup>, J. Karlsson<sup>5</sup>, C. Kelleher<sup>6</sup>, R. Kirkpatrick<sup>11</sup>, M. Kirst<sup>22</sup>, A. Kohler<sup>13</sup>, U. Kalluri<sup>1</sup>, F. Larimer<sup>2</sup>, J. Leebens-Mack<sup>21</sup>, J.-C. Leplé<sup>23</sup>, P. Locascio<sup>2</sup>, Y. Lou<sup>9</sup>, S. Lucas<sup>9</sup>, F. Martin<sup>13</sup>, B. Montanini<sup>13</sup>, C. Napoli<sup>26</sup>, D. R. Nelson<sup>36</sup>, C. Nelson<sup>37</sup>, K. Nieminen<sup>31</sup>, O. Nilsson<sup>12</sup>, V. Pereda<sup>13</sup>, G. Peter<sup>22</sup>, R. Philippe<sup>6</sup>, G. Pilate<sup>23</sup>, A. Poliakov<sup>25</sup>, J. Razumovskaya<sup>2</sup>, P. Richardson<sup>9</sup>, C. Rinaldi<sup>13</sup>, K. Ritland<sup>8</sup>, P. Rouzé<sup>10</sup>, D. Ryaboy<sup>25</sup>, J. Schmutz<sup>28</sup>, J. Schrader<sup>38</sup>, B. Segerman<sup>5</sup>, H. Shin<sup>11</sup>, A. Siddiqui<sup>11</sup>, F. Sterky<sup>39</sup>, A. Terry<sup>9</sup>, C.-J. Tsai<sup>15</sup>, E. Uberbacher<sup>2</sup>, P. Unneberg<sup>39</sup>, J. Vahala<sup>32</sup>, K. Wall<sup>18</sup>, S. Wessler<sup>21</sup>, G. Yang<sup>21</sup>, T. Yin<sup>1</sup>, C. Douglas<sup>7,‡</sup>, M. Marra<sup>11,‡</sup>, G. Sandberg<sup>12,‡</sup>, Y. Van de Peer<sup>10,‡</sup>, D. Rokhsar<sup>9,24,‡</sup>

2006





- 1,000+ natural genotypes planted in three common gardens
- Resequenced to identify 45M SNPs & phenotyped for many years

Population genomics of *Populus trichocarpa* identifies signatures of selection and adaptive trait associations

### Mediation of plant-mycorrhizal interaction by a lectin receptor-like kinase nature plants

A 5-Enolpyruvylshikimate 3-Phosphate Synthase Functions as a Transcriptional Repressor in Populus<sup>[OPEN]</sup>

penetics

Sequencing and Analysis of the Sex Determination Region of *Populus* trichocarpa



Jerry Tusken CBI/ORNL

Wellington Muchero **CBI/ORNL** 

Jay Chen CBI/ORNL Dan Jacobsen **CBI/ORNL** 



Agave REVEILLE1 regulates the onset and release of seasonal dormancy in Populus Plant Physiology<sup>®</sup>

Expression quantitative trait loci mapping identified PtrXB38 as a key hub gene in adventitious root development in *Populus* New



## Outbred diploid P. tremula x P. alba = 717-1B4







<i>the</i> plant journal	<b>S (=)</b> B
The Plant Journal (2023) <b>116.</b> 1003–1017	doi: 10.1111/tpi.16454

SPECIAL ISSUE ARTICLE

### Haplotype-resolved genome assembly of *Populus tremula* × *P. alba* reveals aspen-specific megabase satellite DNA **0 0**

Ran Zhou<sup>1,2,3</sup>, Jerry W Jenkins<sup>4</sup>, Vibing Zeng<sup>2</sup>, Shengqiang Shu<sup>5</sup>, Hosung Jang<sup>2</sup>, Scott A. Harding<sup>1,2,3</sup>, Melissa Williams<sup>4</sup>, Christopher Plott<sup>4</sup>, Kerrie W. Barry<sup>5</sup>, Maxim Koriabine<sup>5</sup>, Mojgan Amirebrahimi<sup>5</sup>, Jayson Talag<sup>6</sup>, Shanmugam Rajasekar<sup>6</sup>, Jane Grimwood<sup>4</sup>, Robert J. Schmitz<sup>2</sup>, R. Kelly Dawe<sup>2,3</sup>, Jeremy Schmutz<sup>4,5</sup>, and Chung-Jui Tsai<sup>1,2,3,\*</sup>





Multiplex knockout of trichome-regulating MYB duplicates in hybrid poplar using a single gRNA

William P. Bewg (1), <sup>1,2,3</sup>, Scott A. Harding (1), <sup>1,2,3</sup>, Nancy L. Engle (1), <sup>4</sup> Brajesh N. Vaidya, <sup>5,†</sup> Ran Zhou (1), <sup>1,2,3</sup>, Jacob Reeves, <sup>6,6</sup> Thomas W. Horn (10), <sup>6</sup> Nirmal Joshee (1), <sup>5</sup> Jerry W. Jenkins (10), <sup>7,8</sup> Shengqiang Shu (10), <sup>8</sup> Kerrie W. Barry (10), <sup>8</sup> Yuko Yoshinaga (10), <sup>8</sup> Jane Grimwood (10), <sup>7,8</sup> Robert J. Schmitz (10), <sup>2</sup> Jeremy Schmutz (10), <sup>7,8</sup> Timothy J. Tschaplinski (10), <sup>4</sup> and Chung-Jui Tsai (10), <sup>1,2,3,6,1</sup>







C.J. Tsai UGA & CBt

# Applying 717

106-22 (9)

0+

106-2



**Early Flower Induction** 



Indeterminate growth after harvest



Morphotypes (architecture/trichome) Chemotypes (bisabolene/cell wall)





**Robin Buell,** Chris Dardick, Wayne Parrott, Bob Schmitz, Patrick Shih, CJ Tsai, Breeanna Urbanowicz



C.J. Tsai UGA & CBI

18



### **Comprehensive Mapping of Long-Range Interactions Reveals Folding Principles of the Human Genome**

Erez Lieberman-Aiden,<sup>1,2,3,4</sup>\* Nynke L. van Berkum,<sup>5</sup>\* Louise Williams,<sup>1</sup> Maxim Imakaev,<sup>2</sup> Tobias Ragoczy,<sup>6,7</sup> Agnes Telling,<sup>6,7</sup> Ido Amit,<sup>1</sup> Bryan R. Lajoie,<sup>5</sup> Peter J. Sabo,<sup>8</sup> Michael O. Dorschner,<sup>8</sup> Richard Sandstrom,<sup>8</sup> Bradley Bernstein,<sup>1,9</sup> M. A. Bender,<sup>10</sup> Mark Groudine,<sup>6,7</sup> Andreas Gnirke,<sup>1</sup> John Stamatoyannopoulos,<sup>8</sup> Leonid A. Mirny,<sup>2,11</sup> Eric S. Lander,<sup>1,12,13</sup>† Job Dekker<sup>5</sup>†



### Dovetail<sup>®</sup> Omni-C<sup>®</sup> Kit

Produce the highest quality genome assembly.

## Haplotype-resolved de novo assembly using phased assembly graphs with hifiasm

Haoyu Cheng, Gregory T. Concepcion, Xiaowen Feng, Haowen Zhang & Heng Li

Nature Methods 18, 170–175 (2021) Cite this article

## Haplotype-resolved assembly of diploid genomes without parental data

Haoyu Cheng, Erich D. Jarvis, Olivier Fedrigo, Klaus-Peter Koepfli, Lara Urban, Neil J. Gemmell & Heng

Nature Biotechnology 40, 1332–1335 (2022) Cite this article



Heng Li Harvard

### Also wrote BWA and Minimap



### Ecotypes & genetic groups





17 - 34 kyrs, ~8k generations

19 - 27 kyrs, last glacial maxima

Demonstrates/proves how selection and evolution act in a complex native grass.

## Today's switchgrass genomes

Release	Size (GB)	Contigs	CN50 (MB)
AP13 V5 CLR	1.10	482	4.3
AP13 V6 H1	1.14	30	55.2
AP13 V6 H2	1.13	33	52.4

4x Upland





4x Lowland<sub>23</sub>

#### JGI CSPs

502977 JBEI – Sequencing Sugarcane BACs 502967-Understanding polyploidy through the generation of the first sugarcane genome sequence - 2017 504319- Genomic Diversity in the Saccharum Complex - 2019



Angelique D'Hont



Karen Aiken





#### **Robert Henry**







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K. S. Aitken<sup>7</sup>, J. Schmutz<sup>14</sup> & A. D'Hont<sup>2,3</sup>

A. L. Healey<sup>127</sup>, O. Garsmeur<sup>2,3</sup>, J. T. Lovell<sup>1,4</sup>, S. Shengquiang<sup>4</sup>, A. Sreedasyam<sup>1</sup>, J. Jenkins<sup>1</sup>,

C. B. Plott<sup>1</sup>, N. Piperidis<sup>5</sup>, N. Pompidor<sup>2,3</sup>, V. Llaca<sup>6</sup>, C. J. Metcalfe<sup>7</sup>, J. Doležel<sup>8</sup>, P. Cápal<sup>8</sup>, J. W. Carlson<sup>4</sup>, J. Y. Hoarau<sup>23,9</sup>, C. Hervouet<sup>2,3</sup>, C. Zini<sup>2,3</sup>, A. Dievart<sup>2,3</sup>, A. Lipzen<sup>4</sup>, M. Williams<sup>1</sup>,

L. B. Boston<sup>1</sup>, J. Webber<sup>1</sup>, K. Keymanesh<sup>4</sup>, S. Tejomurthula<sup>4</sup>, S. Rajasekar<sup>10</sup>, R. Suchecki<sup>11</sup>, A. Furtado<sup>12</sup>, G. May<sup>6</sup>, P. Parakkal<sup>6</sup>, B. A. Simmons<sup>12,13</sup>, K. Barry<sup>4</sup>, R. J. Henry<sup>12,14</sup>, J. Grimwood<sup>1</sup>,

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Published online: 27 March 2024	

Open access



**Brandon James** 

### Sugarcane is complicated



## Multiple sequencing attempts



Release	Size	Contigs	Contig N50
Illumima	5.0 Gb	1,700,000	4.4 Kb
Long read	7.4 Gb	33,222	482 kb
HiFi	9.6 Gb	38,822	10 Mb

### HiFi version

- Better splitting of haplotypes
- ~20X longer contigs
- ~11,000 more genes captured



### **HiC for validation**



### Genetic map (Self1)

ACACGCCTAGGCATCCTACGGTTGTACAATAA ACACGCCTAGGCATCCTACGGTTGTACAATAA

### Single chromosome sort libraries (SCL)





20k genes

ARTICLE 008: 10.1038/s41467-018-05051-5 0F

### A mosaic monoploid reference sequence for the highly complex genome of sugarcane

Olivier Garsmeure<sup>12</sup>, Gaetan Drac<sup>12</sup>, Rudie Antonise<sup>1</sup>, Jane Grimwood<sup>4</sup>, Bernard Potiere<sup>15</sup>, Karen Akiten<sup>6</sup>, Jerry Jenko<sup>45</sup>, Guillaume Marting<sup>12</sup>, Carine Charrone<sup>12</sup>, Catherne Hervouet<sup>12</sup>, Laurett Costel<sup>2</sup>, <sup>1</sup> Nabila Vahiou<sup>43</sup>, Adam Heale<sup>4</sup>, <sup>1</sup>Ovid Sims<sup>4</sup>, Yessei Chenkun<sup>47</sup>, Aninah Seedasyam<sup>4</sup>, Andreg Kian<sup>8</sup>, Agnes Chan<sup>9</sup>, Marie-Anne Van Sluys<sup>10</sup>, <sup>1</sup>Sankhita Swaminathan<sup>4</sup>, Christopher Toam<sup>6</sup>, <sup>1</sup>Hillene Bergle<sup>11</sup>, Blake Simmore<sup>13</sup>, Jean Christophe Glaszmann<sup>13</sup>, Edwin van der Vorsen<sup>3</sup>, Robert Henrye<sup>13</sup>, Jeremy Schmutz<sup>14</sup>, <sup>1</sup>A Angeligue D'Hont<sup>2</sup>. Monoploid 12345678910

R570 1b 1a 1e 1d 1c 1f 5g 5e 5d 5c 5a 5b 5f 59a 2f 2c 2a 2d 2e 2b 2g 3f 3d 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3a 3b 3c 3e 3g 100 0a 0t 010 0710a7d 7b 7c 7a 7a 35 38 0a 3b 3a 3b 3a

### S.spontaneum 1d 1a 1b 1c 2d 2b 2a 2c 3a 3b 3c 3d 4b 4c 4a 4d 5c 5b 5a 5d 6b 6a 6c 6d 7c 7d 7b 7a 8d 8b 8a8d

1

genetics	ARTICLES
Corrected: Publisher Correction	OPEN

### Allele-defined genome of the autopolyploid sugarcane Saccharum spontaneum L.

Jisen Zhang<sup>©1,20+</sup>, Xingtan Zhang<sup>1,20</sup>, Haibao Tang<sup>©1,20</sup>, Qing Zhang<sup>1,20</sup>, Xiuting Hua<sup>1</sup>, Xiaokai Ma<sup>1</sup>, Fan Zhu<sup>2</sup>, Tyler Jones<sup>3</sup>, Xinguang Zhu<sup>4</sup>, John Bowers<sup>5</sup>, Ching Man Wai<sup>4</sup>, Chunfang Zheng<sup>7</sup>, Yan Shi<sup>1</sup>, Shuai Chen', Xiuming Xu', Jingjing Yue', David R. Nelson®, Lixian Huang', Zhen Li', Huimin Xu', Dong Zhou', Yongjun Wang', Weichang Hu', Jishan Lin', Youjin Deng', Neha Pandey<sup>2</sup>, Melina Mancini Dessireé Zerpa<sup>2</sup>, Julie K. Nguyen<sup>2</sup>, Liming Wang<sup>1</sup>, Liang Yu<sup>2</sup>, Yinghui Xin<sup>2</sup>, Liangfa Ge<sup>2</sup>, Jie Arro<sup>2</sup>, Jennifer O. Han<sup>2</sup>, Setu Chakrabarty<sup>2</sup>, Marija Pushko<sup>2</sup>, Wenping Zhang<sup>1</sup>, Yanhong Ma<sup>1</sup>, Panpan Ma<sup>1</sup>, Mingju Lv4, Faming Chen9, Guangyong Zheng9, Jingsheng Xu1, Zhenhui Yang1, Fang Deng1, Xuequn Chen<sup>1</sup>, Zhenyang Liao<sup>1</sup>, Xunxiao Zhang<sup>1</sup>, Zhicong Lin<sup>1</sup>, Hai Lin<sup>1</sup>, Hansong Yan<sup>1</sup>, Zheng Kuang Weimin Zhong', Pingping Liang', Guofeng Wang', Yuan Yuan', Jiaxian Shi', Jinxiang Hou', Jingxian Lin' Jingjing Jin<sup>10</sup>, Peijian Cao<sup>10</sup>, Qiaochu Shen<sup>1</sup>, Qing Jiang<sup>1</sup>, Ping Zhou<sup>1</sup>, Yaying Ma<sup>1</sup>, Xiaodan Zhang<sup>1</sup>, Rongrong Xu', Juan Liu', Yongmei Zhou', Haifeng Jia', Qing Ma', Rui Qi', Zhiliang Zhang', Jingping Fang', Hongkun Fang', Jinjin Song', Mengjuan Wang', Guangrui Dong', Gang Wang', Zheng Chen', Teng Ma', Hong Liu<sup>1</sup>, Singha R. Dhungana<sup>9</sup>, Sarah E. Huss<sup>2</sup>, Xiping Yang<sup>12</sup>, Anupma Sharma<sup>11</sup>, Jhon H. Trujillo<sup>14</sup>, Maria C. Martinez<sup>94</sup>, Matthew Hudson<sup>95</sup>, John J. Riascos<sup>94</sup>, Mary Schuler<sup>2</sup>, Li-Qing Chen<sup>2</sup>, David M. Braun", Lei Li<sup>1</sup>, Qingyi Yu<sup>013</sup>, Jianping Wang<sup>0132</sup>, Kai Wang<sup>1</sup>, Michael C. Schatz<sup>16</sup>, David Heckerman<sup>90</sup>, Marie-Anne Van Sluvs<sup>18</sup>, Glaucia Mendes Souza<sup>919</sup>, Paul H. Moore<sup>3</sup> David Sankoff', Robert VanBuren\*, Andrew H. Paterson\*, Chifumi Nagai\* and Ray Ming

Chromosomes scaled by gene rank order

2 3 4 5 6 7 8 9 10

Primary Path 5.1 Gb Genome (Primary Path) 844 Contigs (N50: 15Mb) 68 Chromosomes



HUDSONALPHA

S.bicolor

Today: We can do this again with just HiFi and Omni-C

## Brown Rust Resistance (Bru1) - Puccinia melanocephala



**R570 Selfed offspring** 



- Before 1980, brown rust caused ~50% yield losses
- From R570, the first resistant cultivar, PCR markers were developed
- Single-copy haplotype-specific insertion that does not recombine
- Candidate gene ended up a two gene, kinase-pseudokinase ٠ complex

## Terpenes in conifer genomes CSP 503037





Jorg Bohlmann UBC

Jay Keasling LBNL/JBEI

**Elzabeth Sattely** Standford





Taxus baccata English Yew 10GB x 2



Western Red Cedar



Jeffrey's pine



**Octane Scale** 

Journal of Essential Oil Research Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/tieo20

Alkanes and Terpenes in Wood and Leaves of Pinus jeffreyi and P. sabiniana Robert P. Adams <sup>a</sup> & Jessica W. Wright <sup>b</sup>

#### \* Images Wikipedia

## Comparative genomics in large genomes



Pinus jeffreyi: 25.2 GBs (63x larger than poplar)

## Great challenge in plant genomics

Genome	No. genes	% GGF
A. thaliana Araport11	27 655	91.33
B. distachyon	34 310	72.56
C. reinhardtii	17 741	43.08
E. grandis	36 349	79.74
G. max	52 872	80.37
K. fedtschenkoi	30 964	82.01
M. truncatula	50 894	67.94
P. hallii var. filipes	33 805	72.65
P. hallii var. hallii	33 263	73.36
P. patens	32 926	55.44
P. trichocarpa	34 699	82.31
P. virgatum	80 278	69.2
S. bicolor	34 129	71.51
S. bicolor Rio	35 490	69.16
S. fallax	25 100	78.31
<u>S. italica</u>	34 584	77
S. viridis	38 334	70.43
L. albus	38 258	78.17

- What do these genes do?
- What effect does variation in these genes have on our traits of interest?
- What about polyploids?

Research

**BdERECTA** controls vasculature patterning

and phloem-xylem organization in

Brachypodium distachyon



Debbie Laudencia-Chingcuanco, USDA CSP #1670

mental Botany, Vol. 67, No. 1 pp. 227-237, 2016

i:10.1093/bds/erv446 Advance Access publication 3 October 2015

Grasses use an alternatively wired bHLH transcription

Michael T. Raissig<sup>a,1</sup>, Emily Abrash<sup>a,1</sup>, Akhila Bettadapur<sup>b</sup>, John P. Vogel<sup>c</sup>, and Dominique C. Bergmann<sup>a,b,2</sup>

**Open Access** 

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Original Article 🙆 Open Access

BMC Plant Biology KARRIKIN INSENSITIVE2 regulates leaf development, root system architecture and arbuscularmycorrhizal symbiosis in Brachypodium distachyon



#### Mobile MUTE specifies subsidiary cells to build physiologically improved grass stomata

Michael T. Raissig,<sup>1</sup>\* Juliana L. Matos,<sup>1</sup> M. Ximena Anleu Gil,<sup>2</sup> Ari Kornfeld,<sup>3</sup> Akhila Bettadapur,<sup>2</sup> Emily Abrash,<sup>1</sup> Hannah R. Allison,<sup>1</sup> Grayson Badgley,<sup>3</sup> John P. Vogel,<sup>4</sup> Joseph A. Berry,<sup>3</sup> Dominique C. Bergmann<sup>1,2</sup>\*

Published online 1 August 2023

#### Nucleic Acids Research, 2023, Vol. 51, No. 16 8383-8401 https://doi.org/10.1093/narlgkad616

JGI Plant Gene Atlas: an updateable transcriptome resource to improve functional gene descriptions across the plant kingdom

RESEARCH PAPER

New Phytolog

Mutation in *Brachypodium* caffeic acid *O*-methyltransferase 6 alters stem and grain lignins and improves straw saccharification without deteriorating grain quality

Mutations in the predicted DNA polymerase subunit POLD3

result in more rapid flowering of Brachypodium distachyon

RESEARCH

DOI: 10.1038/s41467-017-02292-8

#### Extensive gene content variation in the Brachypodium distachyon pan-genome correlates with population structure

OPEN

Sean P. Gordon<sup>1</sup>, Bruno Contreras-Moreira <sup>2,3,4</sup>, Daniel P. Woods<sup>5,6</sup>, David L. Des Marais <sup>7,17</sup>, Diane Burgess<sup>8</sup>, Shengqiang Shu<sup>1</sup>, Christoph Stritt<sup>9</sup>, Anne C. Roulin<sup>9</sup>, Wendy Schackwitz<sup>1</sup>, Ludmila Tyler<sup>10</sup>, Joel Martin 6 <sup>1</sup>, Anna Lipzen<sup>1</sup>, Niklas Dochy <sup>11</sup>, Jeremy Phillips<sup>1</sup>, Kerrie Barry<sup>1</sup>, Koen Geuten <sup>11</sup>, Hikmet Budak o 12, Thomas E. Juenger 13, Richard Amasino o 5,6, Ana L. Caicedo 10, David Goodstein 1, Patrick Davidson<sup>1</sup>, Luis A. J. Muro<sup>14</sup>, Melania Figueroa<sup>15</sup>, Michael Freeling<sup>8</sup>, Pilar Catalan<sup>4,16</sup> & John P. Vogel 1,8



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Shell (R)

(10,060)

Shell (N)

(7,135)/Softcore

(7, 283)

Core

(13, 408)

nature COMMUNICATIONS



### Microbe references

Koonin EV, Makarova KS, Wolf YI. Evolution of Microbial Genomics: Conceptual Shifts over a Quarter Century. Trends Microbiol. 2021 Jul;29(7):582-592.

ARTICLE

## Switchgrass pangenome



Pangenomes allow us to capture diversity in structural variants that we can't easily identify with short read sequencing

## Switchgrass gene cluster variation



## A switchgrass pangenome graph



Fig. 1 | References are well represented in final pangenome graph



Chr01K

Fig. 3 | Test locus: variation in ABO3 (AP13 HAP1 Chr08K:838322-839442)





Graph construction: minigraph-cactus Read alignment to graph: vg giraffe Viz via subway maps: sequenceTubeMap

Hickey, G., Monlong, J., Ebler, J. *et al.* Pangenome graph construction from genome alignments with Minigraph-Cactus. *Nat Biotechnol* (2025).

#### On average,

89%

of each input reference genome is retained in the final graph

80%

ò

of clipped sequences are annotated repeats

### How much diversity do we cover (10 genomes, 20 haplotypes)?

Gulf subpopulation



Lowland ecotype (n = 105) Coastal (n = 63)Coastal (n = 73)Upland (n = 117) Upland (n = 196)Gulf Midwest Atlantic 5B **−−−** coverage ≥ 1, quorum ≥ 0%,  $k_1 X^{\gamma}$  with  $k_1 \neq 784.2$  M,  $\gamma = 0.598$ ) coverage  $\geq 1$ , quorum  $\geq 0\%$ coverage  $\geq 2$ , guorum  $\geq 0\%$ coverage  $\geq 1$ , quorum  $\geq 50\%$ 4Bcoverage ≥ 1, quorum ≥ 100%3B #bps 2B 1B 0 δ  $\checkmark$ 5 9  $\overline{}$ \$ 0 20  $\sim$ 13 14 14 26 ~ 28 20 0 77 29 samples

Atlantic subpopulation

Midwest subpopulation

## Pangenome graphs in simpler systems

### Camelina (12)





CSP2020 505836





JOINT GENOME INSTITUTE



John Mullet Texas A&M

BILL&MELINDA GATES foundation



ດດາວາ



Geoff Morris Nadia Shakoor Colorado State Donald Danforth

🖉 CABB



Bird et al. bioRxiv 2024.08.13.607619; doi: https://doi.org/10.1101/2024.08.13.607619

BERSS 507613

## Example gene

\_ . . . . . .





Striga – witchweed \*

## LGS1 in pangenome



## Mapping LGS1 variants across Africa



# Field phenotyping













Nadia Shakoor Donald Danforth

### Leaf Water Management

## Miscanthus : embracing complexity



Kankshita Swaminathan CABBI, HudsonAlpha



**Erik Sacks** 



200 Mbp

Chromosomes scaled by physical position



Sorghum

4x –Hap1

4x –Hap2

**Unpublished Data** 42

## Goals of customization for perennial feedstocks



- Improve <u>field traits</u>: high biomass, improved germination & maturation, lodging resistance, leaf drop, flowering time, reduced fertilizer
- Improve <u>breeding traits</u>: shattering, self compatibility, sterility
- Improve <u>quality/processing traits</u>: fiber length, ash, chemical composition, fiber/lignan rations, reduce internodes, low pith
- Make into chemical factories

## Editing in complex plants







Kankshita Swaminathan Tony Trieu Team Swaminathan



wild-type edited line M. sinensis PMS-014



Pradeep Hirannaiah



Joshua Stanley NSF-BRIDGES

Unpublished



In tetraploids and hexaploid production genotype

lw1

WT

## Engineering Miscanthus to produce vegetative lipids



GCB Bioenergy, Volume: 13, Issue: 10, Pages: 1610-1623, First published: 26 July 2021, DOI: (10.1111/gcbb.12883)

- "Push" Photosynthetic carbon into fatty acid biosynthesis
- Wrinkled 1 transcription factor OE
- "Pull" Pull fatty acids into TAG
- DGAT diacyglycerol acyltransferase OE "Protect" - Block TAG breakdown
- Oleosin OE, lipase suppression

Unpublished data



Trieu

## Our legacy – JGI impact on plant genomes

Phytozome 🖉 13

THE PLANT GENOMICS RESOURCE

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Velcome to Phytozome	Phytozome <b>v</b>		
Overview	Release Notes	s News	
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Sorghum bicolor	v5.1	sweet sorghum	Feb 1, 2023
Sorahum bicolor	Wray v1.1	sweet sorghum	Feb 1, 2023
oorginam brootor		Wray	
Setaria viridis v4	1	Wray green foxtail millet	Feb 1, 2023
Setaria viridis v4. Gossypium hirsu	1 tum Coker	Wray green foxtail millet upland cotton	Feb 1, 2023 Feb 1, 2023
Setaria viridis v4. Gossypium hirsu Gossypium hirsu	1 tum Coker tum DeltaPearl	Wray green foxtail millet upland cotton upland cotton	Feb 1, 2023 Feb 1, 2023 Feb 1, 2023
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ome, the Plant Comparative Genomics portal of the Department of Energy's enome Institute, provides JGI users and the broader plant science nity a hub for accessing, visualizing and analyzing JGI-sequenced plant es, as well as selected genomes and datasets that have been sequenced ere. By integrating this large collection of plant genomes into a single and performing comprehensive and uniform annotation and analyses, ome facilitates accurate and insightful comparative genomics studies.

lease v13, Phytozome hosts 411 assembled and annotated genomes, from 174 Archaeplastida species, and contains the 54 Brachypodium distachyon lines from the BrachyPan pan-genome study, the 20 species included in the Brassicales Man Alignment Project, and 8 cownea (Vigna unguiculata) genomes

find genes by keyword search by BLAST get standard data files build cu Panicum virgatum var. DAC6 HAP2 v1.1 ① RST JGI Panicum virgatum var. VS16 HAP1 v1.1 ① RST JGI Panicum virgatum var. VS16 HAP2 v1.1 ① RST JGI Panicum virgatum var. WBC HAP1 v1.1 ① RST JGI Panicum virgatum var. WBC HAP2 v1.1 ① RST JGI Paspalum vaginatum v3.1 ① UNRST JGI Saccharum officinarum x spontaneum R570 v2.1 ① RST JGI Setaria italica v2.2 ① UNRST GeneAtlas JGI Setaria viridis v1.1 ① UNRST GeneAtlas JGI Setaria viridis v2.1 ① UNRST GeneAtlas JGI Setaria viridis v4.1 ① RST JGI Urochloa fusca v1.1 ② RST JGI JURST GeneAtlas JGI Setaria viridis v4.1 ② RST JGI Zea mays RefGen_V4 ③ UNRST EXT Zea mays B84 v1.2 ③ UNRST JGI Zea mays LH145 v1.2 ④ UNRST JGI Zea mays NKH8431 v1.2 ① UNRST JGI Zea mays PH207 v1.1 ④ UNRST JGI	Choose genomes by sel	ecting from tree or typ	pe genus/species/commo	1 genom
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<ul> <li>Paspalum vaginatum v3.1 () UNRST JGI</li> <li>Saccharum officinarum x spontaneum R570 v2.1 () RST JGI</li> <li>Setaria italica v2.2 () UNRST GeneAtlas JGI</li> <li>Setaria viridis v1.1 () UNRST GeneAtlas JGI</li> <li>Setaria viridis v4.1 () RST JGI</li> <li>Urochloa fusca v1.1 () RST JGI</li> <li>Urochloa fusca v1.1 () RST JGI</li> <li>Zea mays RefGen_V4 () UNRST EXT</li> <li>Zea mays B84 v1.2 () UNRST JGI</li> <li>Zea mays LH145 v1.2 () UNRST JGI</li> <li>Zea mays NKH8431 v1.2 () UNRST JGI</li> <li>Zea mays PH207 v1.1 () UNRST EXT</li> </ul>	— Panicu	ım virgatum var. WBC H	AP2 v1.1 () RST JGI	
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<ul> <li>Setaria italica v2.2 (i) UNRST GeneAtlas JGI</li> <li>Setaria viridis v1.1 (i) UNRST GeneAtlas JGI</li> <li>Setaria viridis v2.1 (i) UNRST GeneAtlas JGI</li> <li>Setaria viridis v4.1 (i) RST JGI</li> <li>Urochloa fusca v1.1 (i) RST JGI</li> <li>Urochloa fusca v1.1 (i) RST JGI</li> <li>Zea mays RefGen_V4 (i) UNRST EXT</li> <li>Zea mays Zm-B73-REFERENCE-NAM-5.0.55 (i) UNRST EXT</li> <li>Zea mays B84 v1.2 (i) UNRST JGI</li> <li>Zea mays LH145 v1.2 (i) UNRST JGI</li> <li>Zea mays NKH8431 v1.2 (i) UNRST JGI</li> <li>Zea mays PH207 v1.1 (i) UNRST EXT</li> </ul>	Sacchi	arum officinarum x spor	ntaneum R570 v2.1 🛈 📧 🕻	JGI
<ul> <li>Setaria viridis v1.1 (i) UNRST GeneAtlas JGI</li> <li>Setaria viridis v2.1 (i) UNRST GeneAtlas JGI</li> <li>Setaria viridis v4.1 (i) RST JGI</li> <li>Urochloa fusca v1.1 (i) RST JGI</li> <li>Urochloa fusca v1.1 (i) RST JGI</li> <li>Zea mays RefGen_V4 (i) UNRST EXT</li> <li>Zea mays Zm-B73-REFERENCE-NAM-5.0.55 (i) UNRST EXT</li> <li>Zea mays B84 v1.2 (i) UNRST JGI</li> <li>Zea mays LH145 v1.2 (i) UNRST JGI</li> <li>Zea mays NKH8431 v1.2 (i) UNRST JGI</li> <li>Zea mays PH207 v1.1 (i) UNRST EXT</li> </ul>	— Setaria	a italica v2.2 (i) UNRST	GeneAtlas JGI	
<ul> <li>Setaria viridis v2.1 ① UNRST GeneAtlas JGI</li> <li>Setaria viridis v4.1 ③ RST JGI</li> <li>Urochloa fusca v1.1 ④ RST JGI</li> <li>Urochloa fusca v1.1 ④ RST JGI</li> <li>Zea mays RefGen_V4 ④ UNRST EXT</li> <li>Zea mays Zm-B73-REFERENCE-NAM-5.0.55 ④ UNRST EXT</li> <li>Zea mays B84 v1.2 ④ UNRST JGI</li> <li>Zea mays LH145 v1.2 ④ UNRST JGI</li> <li>Zea mays NKH8431 v1.2 ④ UNRST JGI</li> <li>Zea mays PH207 v1.1 ④ UNRST EXT</li> </ul>	— Setaria	a viridis v1.1 (i) UNRST	GeneAtlas JGI	
<ul> <li>Setaria viridis v4.1 (i) RST JGI</li> <li>Urochloa fusca v1.1 (i) RST JGI</li> <li>Maize</li> <li>Zea mays RefGen_V4 (i) UNRST EXT</li> <li>Zea mays Zm-B73-REFERENCE-NAM-5.0.55 (i) UNRST EXT</li> <li>Zea mays B84 v1.2 (i) UNRST JGI</li> <li>Zea mays LH145 v1.2 (i) UNRST JGI</li> <li>Zea mays NKH8431 v1.2 (i) UNRST JGI</li> <li>Zea mays PH207 v1.1 (i) UNRST EXT</li> </ul>	Setaria	a viridis v2.1 (i) UNRST	GeneAtlas JGI	
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<ul> <li>Maize</li> <li>Zea mays RefGen_V4 ① UNRST EXT</li> <li>Zea mays Zm-B73-REFERENCE-NAM-5.0.55 ① UNRST EXT</li> <li>Zea mays B84 v1.2 ① UNRST JGI</li> <li>Zea mays LH145 v1.2 ① UNRST JGI</li> <li>Zea mays NKH8431 v1.2 ① UNRST JGI</li> <li>Zea mays PH207 v1.1 ② UNRST EXT</li> </ul>	— Urochl	oa fusca v1.1 🛈 📧	JGI	
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— Zea mays LH145 v1.2 ① UNRST JGI — Zea mays NKH8431 v1.2 ① UNRST JGI — Zea mays PH207 v1.1 ② UNRST EXT	— Zea	mays B84 v1.2 🛈 UNR	IST JGI	
— Zea mays NKH8431 v1.2 ① UNRST JGI — Zea mays PH207 v1.1 ③ UNRST EXT	— Zea	mays LH145 v1.2 🛈 🕻	JNRST JGI	
Zea mays PH207 v1.1 () UNRST EXT	— Zea	mays NKH8431 v1.2 🤇	UNRST JGI	
	— Zea	mays PH207 v1.1 🛈 🛛	JNRSTEXT	
	— Zea	mays PHB47 v1.2 🛈	UNRST JGI	

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2024

JGI Home



•	19.4	gallons	gasoline
•	12.5	gallons	diesel/home heating oil
٠	4.4	gallons	kerosene-type jet fuel
•	2.1	gallons	coke
٠	1.6	gallons	still gas
•	1.5	gallons	liquefied refinery gases
•	0.9	gallons	heavy, residual fuel oils
•	0.8	gallon	asphalt and road oil
•	0.5	gallon	petrochemical feedstocks
•	0.4	gallon	lubricants
•	0.4	gallon	other



42-gallon barrel of crude

https://www.lubesngreases.com/magazine/28\_8/gasoline-and-engineoil-the-whole-story/





These are only examples of discoveries in plant genomics- 1000's of pathways and variant for traits of interest are needed to customize and build genotypes for end uses.

### A Unified Data Infrastructure for Biological and Environmental Research

Report from the BER Advisory Committee



**Overcoming Barriers in Plant Transformation** *A Focus on Bioenergy Crops* 



BER is uniquely positioned to accelerate these effort with a combination of climate modeling for global biogeochemistry, BRCs for deconstruction/customization of feedstocks, and the combined efforts of JGI, national and academic labs to acquire and develop the biological, environmental knowledge to make this possible.







**Biological and Environmental Research** 

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